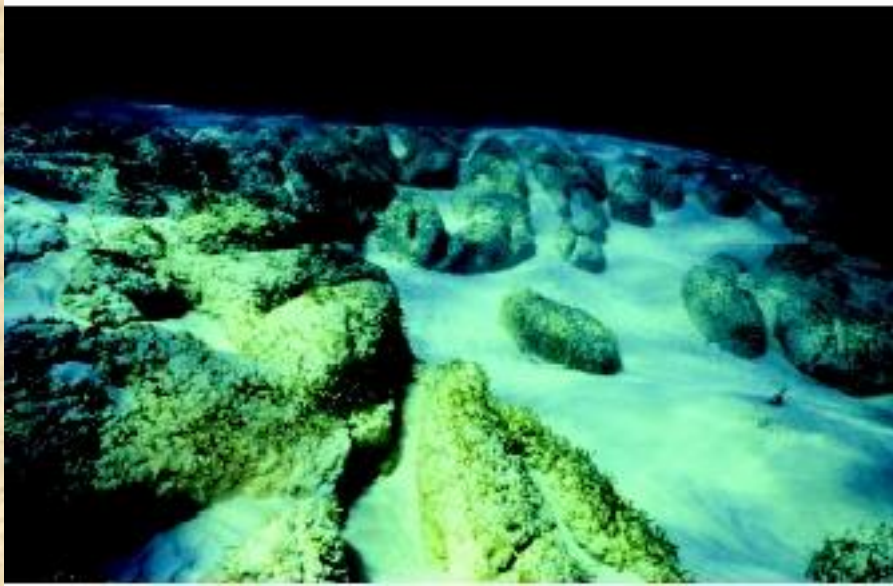


# Factors Influencing Cyanobacteria Blooms in Farmington Bay, Great Salt Lake, Utah.

Brad Marden Leland Myers Theron Miller David Richards

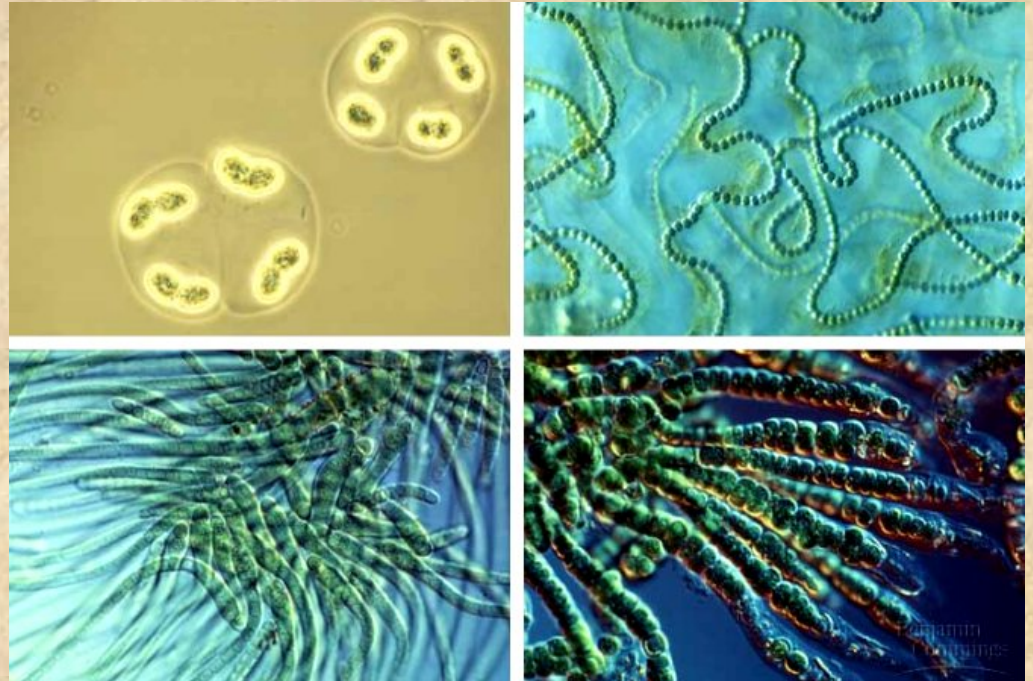






The ability of cyanobacteria to perform oxygenic photosynthesis is thought to have converted the early reducing atmosphere into an oxidizing one, which dramatically changed the composition of life forms on Earth by stimulating biodiversity and leading to the near-extinction of oxygen-intolerant organisms.

Cyanobacteria are arguably the most successful group of microorganisms on earth. They are the most genetically diverse; they occupy a broad range of habitats across all latitudes, widespread in freshwater, marine and terrestrial ecosystems, and they are found in the most extreme niches such as hot springs, salt works, and hypersaline bays.





# GSL Stromatolites

Cover about 10% of Lake bottom

Calcium/Magnesium Cyanobacteria  
Deposits

Primary substrate for Brine Fly  
Pupae life stage



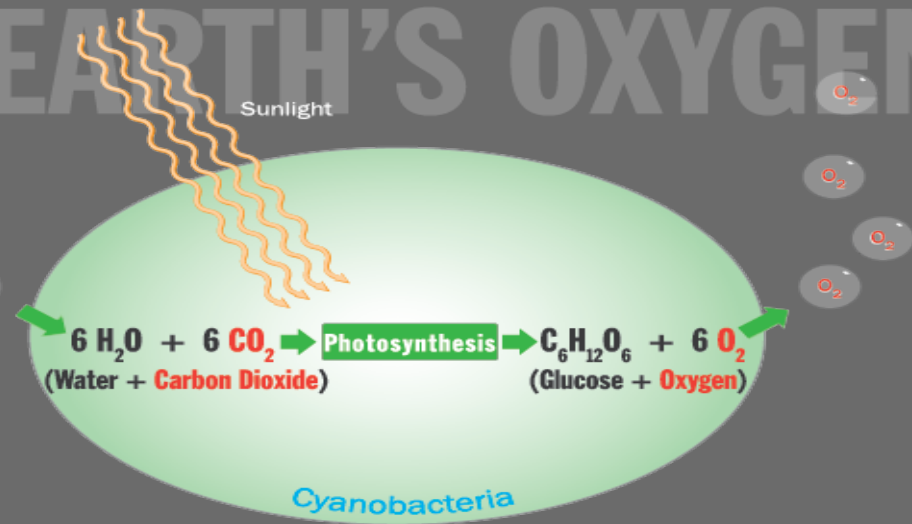


# 1/4 OF EARTH'S OXYGEN

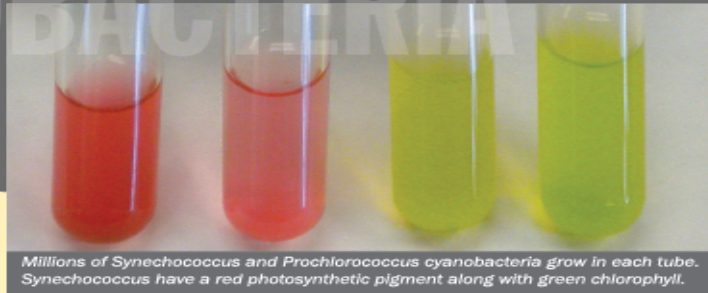
## CYANOBACTERIA — WORLD'S SMALLEST OXYGEN PRODUCERS

Cyanobacteria take in carbon dioxide and release oxygen during photosynthesis.

Evolving about 2.5 billion years ago, cyanobacteria were Earth's first photosynthesizers, the first living source of oxygen on earth.

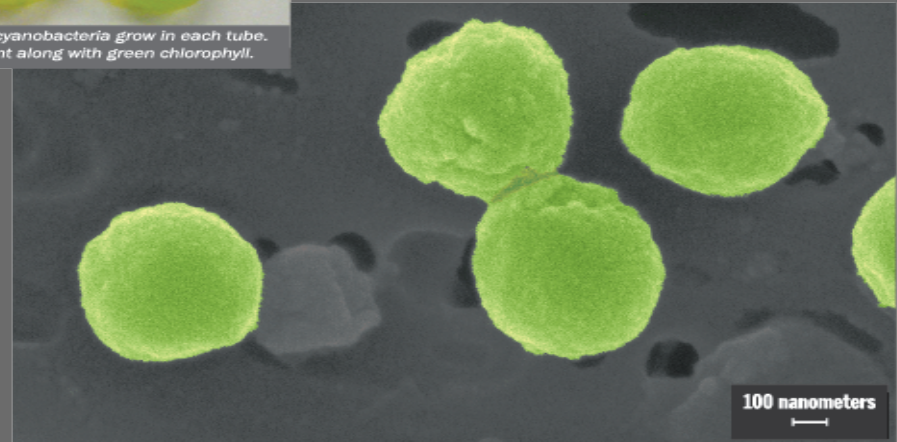
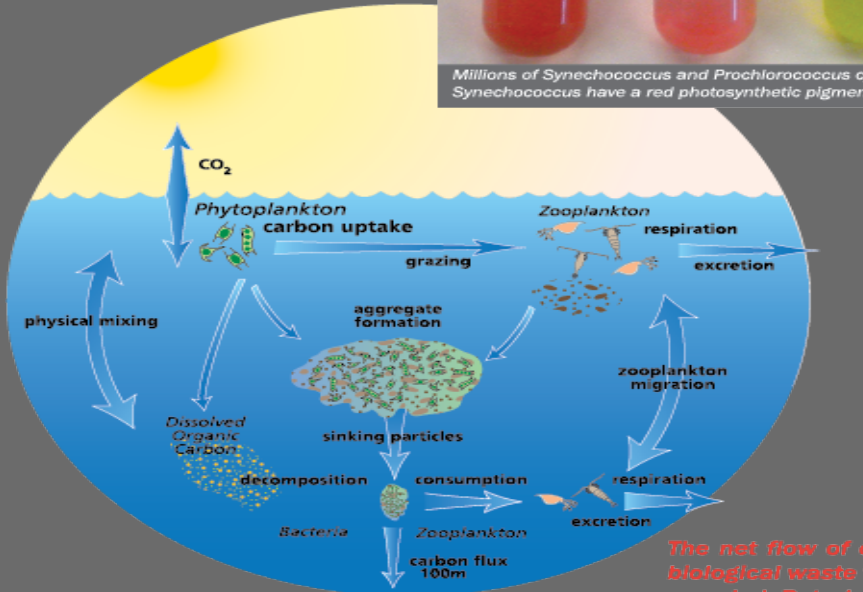


# CYANOBACTERIA



« ... the most important microbe you've never heard of. »

Joe Palca, National Public Radio  
 August 2008, 20th anniversary  
 of the discovery of *Prochlorococcus*



False-color scanning electron micrograph of *Prochlorococcus* bacteria

The net flow of carbon is into the ocean, because of biological waste that falls to the ocean floor and is not recycled. Petroleum extraction reverses this process.





**Notice**  
An algae bloom has made this area potentially unsafe for water contact. Avoid direct contact with visible surface scum.





# Hepatotoxins (Microcystin/ Nodularian)

- Fitzgeorge et al. (1994) demonstrated that microcystin [and other hepatotoxins] toxicity is cumulative: a single oral dose resulted in no increase in liver weight (which is a measure of liver damage), whereas the same dose applied daily over seven days caused an increase in liver weight of 84% and thus had the same effect as a single oral dose 16 times as large.



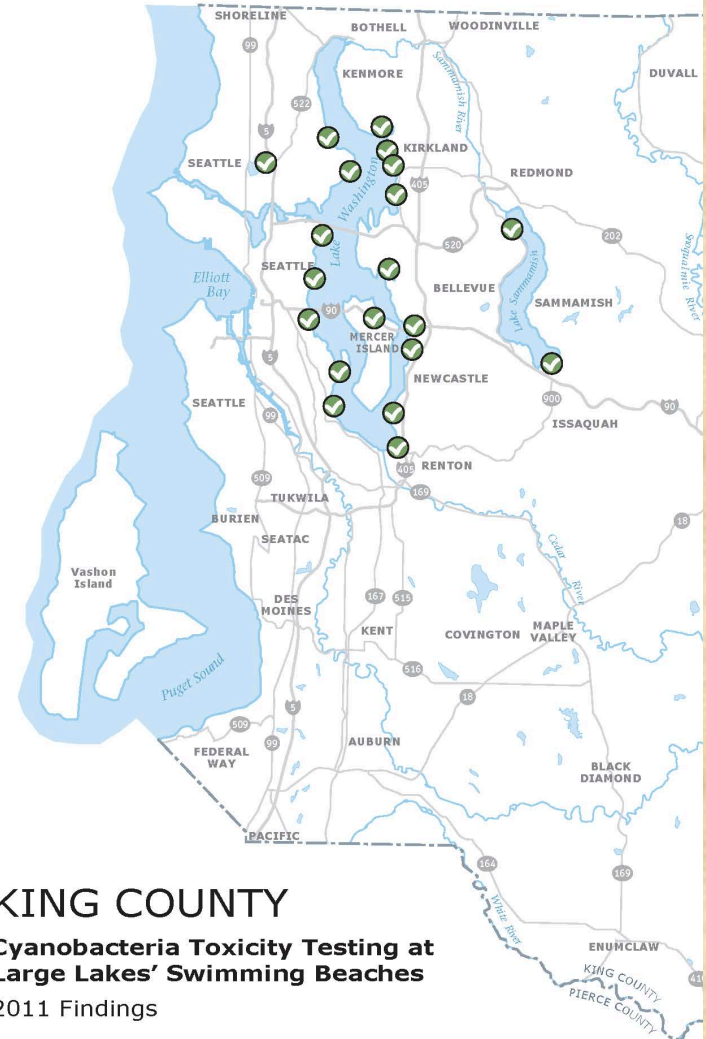


# Anatoxins

Saxitoxins and anatoxin-a(s) are among the most neurotoxic substances known. However, evidence is accumulating that in lakes and rivers they do not occur as frequently as microcystins. This applies especially to anatoxin-a: to date, it has been found only in a small number of *Anabaena* blooms in North America. Furthermore, concentrations even of these highly toxic substances in scums will scarcely reach levels acutely neurotoxic to a human ingesting a mouthful. In contrast, neurotoxicity may be experienced by livestock that drink many litres of contaminated water and pets—especially dogs—that gather scum material in their fur and ingest it through grooming with the tongue.

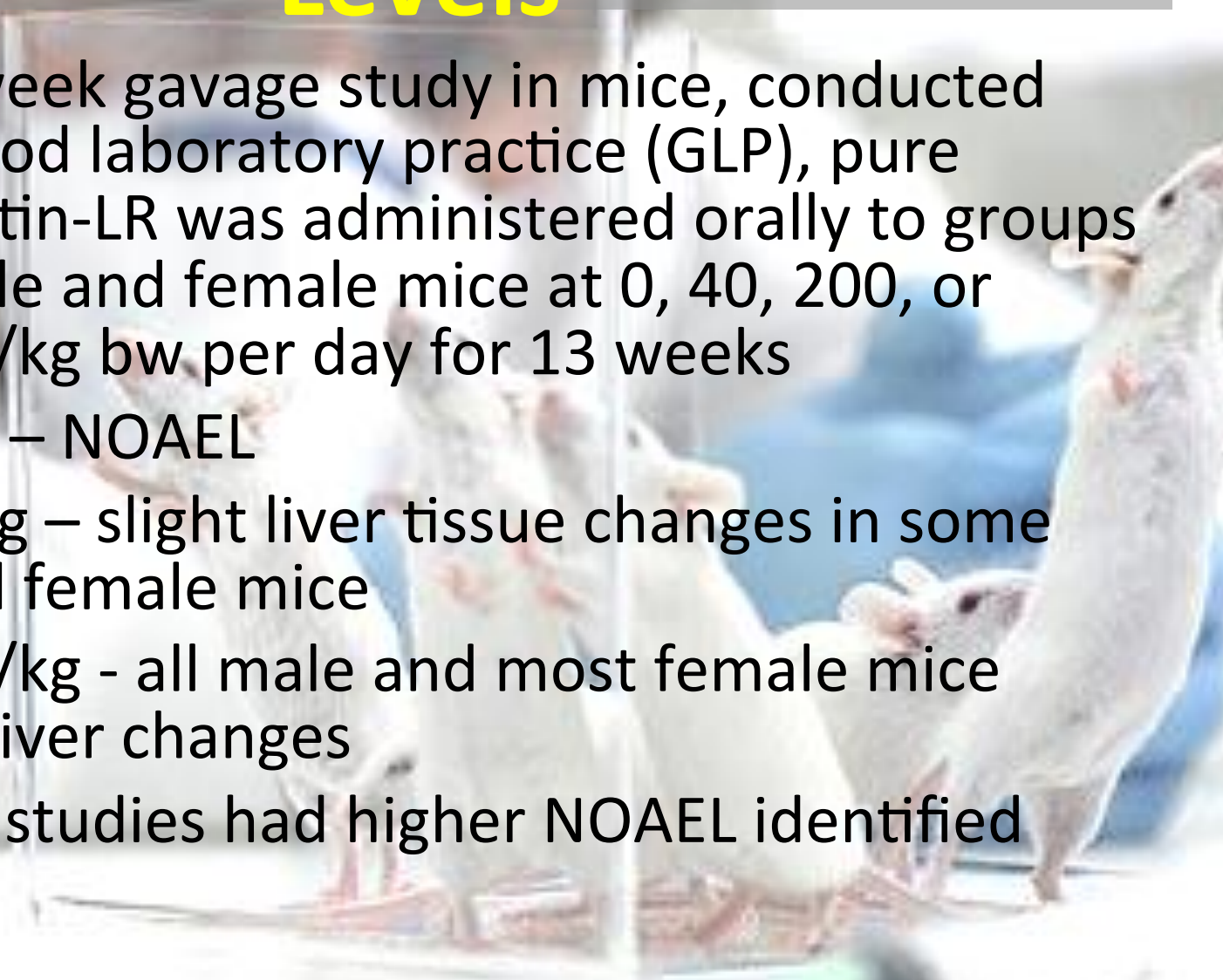


# What is too much Cyanotoxins ???





# Development of Warning Levels

- In a 13-week gavage study in mice, conducted under good laboratory practice (GLP), pure microcystin-LR was administered orally to groups of 15 male and female mice at 0, 40, 200, or 1,000  $\mu\text{g}/\text{kg}$  bw per day for 13 weeks
  - 40  $\mu\text{g}/\text{kg}$  – NOAEL
  - 200  $\mu\text{g}/\text{kg}$  – slight liver tissue changes in some male and female mice
  - 1,000  $\mu\text{g}/\text{kg}$  - all male and most female mice showed liver changes
  - All other studies had higher NOAEL identified
- 
- A photograph of several white mice in a laboratory setting. The mice are in various poses, some standing on their hind legs, others sitting or lying down. The background is slightly blurred, showing what appears to be a laboratory environment with some equipment and a person's legs in blue pants.

# Development of Warning Levels

$$\text{Guideline Value} = \frac{(\text{Tolerable Daily Intake}) (\text{Body Wt.} - \text{Kg}) (\text{Proportion H}_2\text{O Contaminated})}{\text{Liters of Water}}$$

- Assumed a typical individual weight of 60 Kg (132 lbs)
- Liters of water drunk daily – 2 Liters
- Proportion from Contaminated Water Source – 80%
- Safe Tolerable Daily Concentration – 0.04  $\mu\text{g}/\text{kg}$  bw per day

40  $\mu\text{g}/\text{kg}$  bw per day

uncertainty factor of 1,000

(10 for intra-species variability, 10 for inter-species variability and 10 for database limitations)

**Drinking Water – 1.0  $\mu\text{g}/\text{L}$**

**Recreation Water – 20  $\mu\text{g}/\text{L}$  (assumes 200 ml Ingested)**



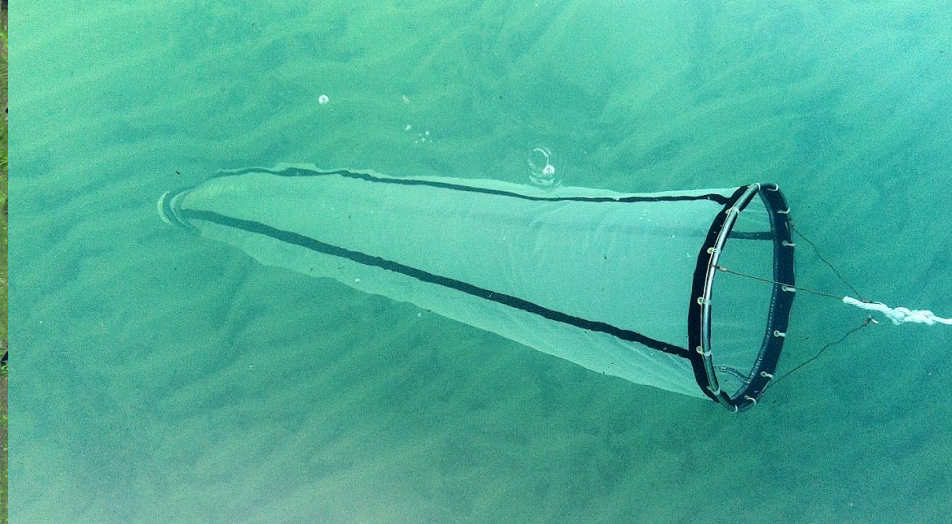
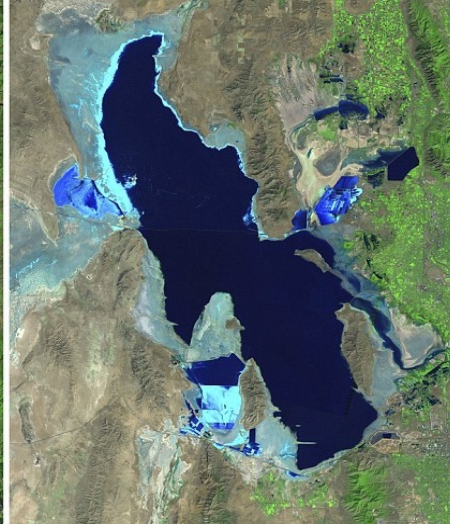
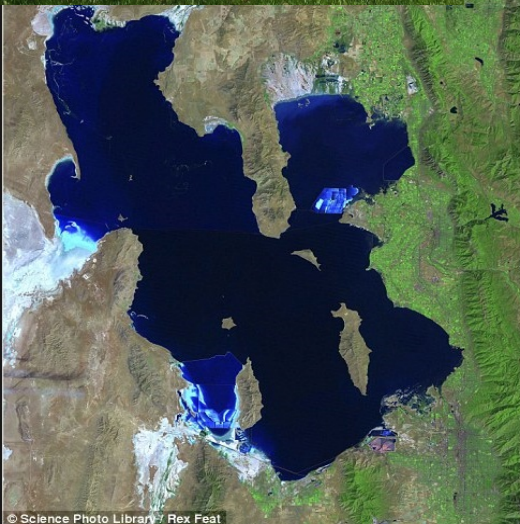
# ***Toledo Experience***





# FBAY Research Program

Cooperation with GSLC





The background of the slide features a scenic landscape with a calm body of water in the foreground, a range of snow-capped mountains in the middle ground, and a clear blue sky above. The overall tone is light and natural.

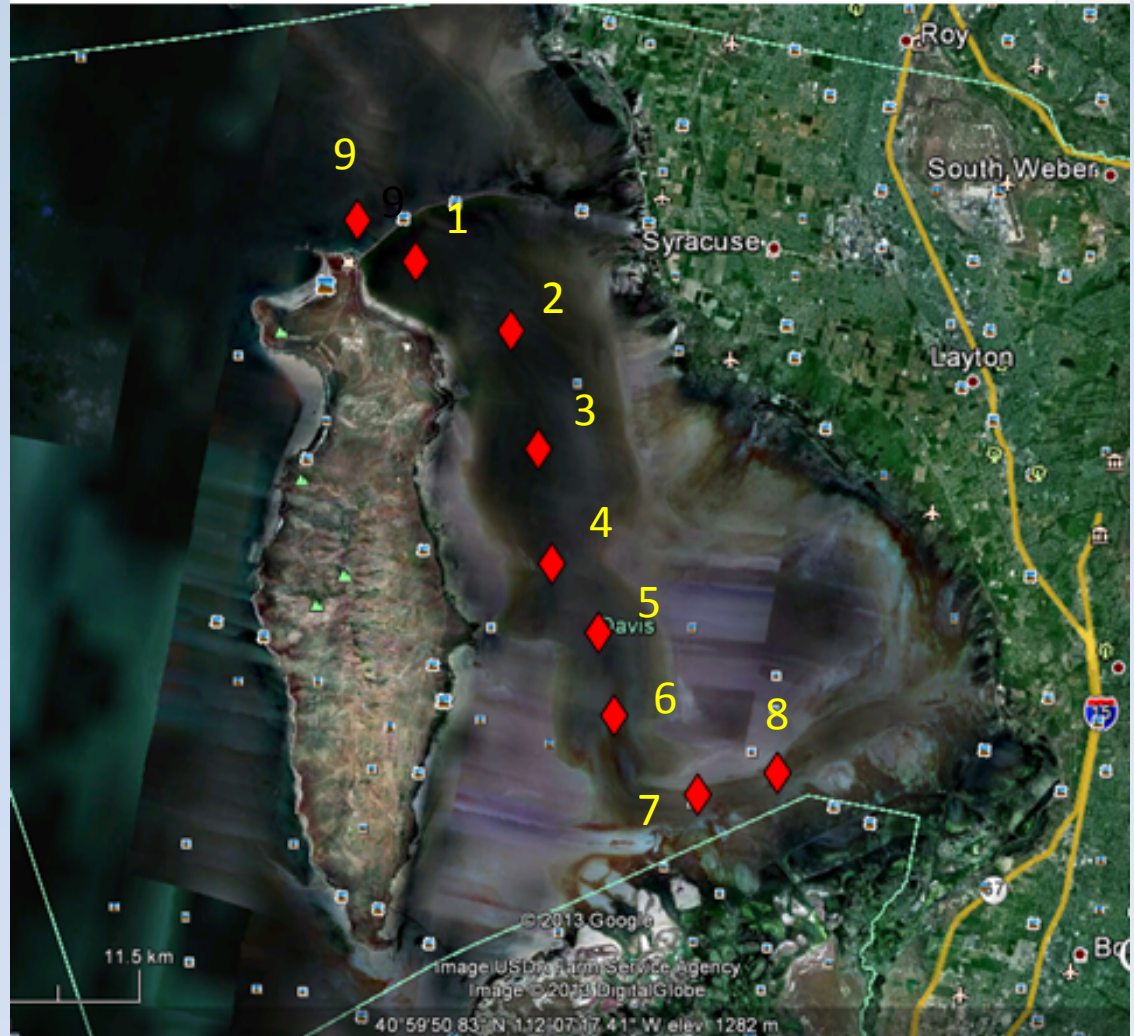
# Project Goals of a Multi-year Investigation of Cyanobacteria Blooms in Farmington Bay

- **Collect a systematic record of the spatial and temporal changes in the biotic community and abiotic characteristics of Farmington Bay from March through November.**
- **Correlate changes in the algal population size and structure and with changes in abiotic factors such as nutrient concentrations, salinity, temperature, and water transparency .**
- **Correlate changes in the macroinvertebrate population composition and abundance, with changes in abiotic factors such as nutrient concentrations, salinity, and temperature, as well as with algal population size and structure, and cyanotoxin concentration.**
- **Identify the linkage between cyanobacteria blooms and cyanotoxin production in Farmington Bay.**
- **Correlate isotopic  $^{15}\text{N}$  and  $^{13}\text{C}$  levels in macroinvertebrates and algae with nutrient sources.**

# Sampling Locations and Totals

## FBAY Research Program

- 14 Transect Surveys
- February to November
- 23 Single Point Collections
- January to December





# Program Schedule

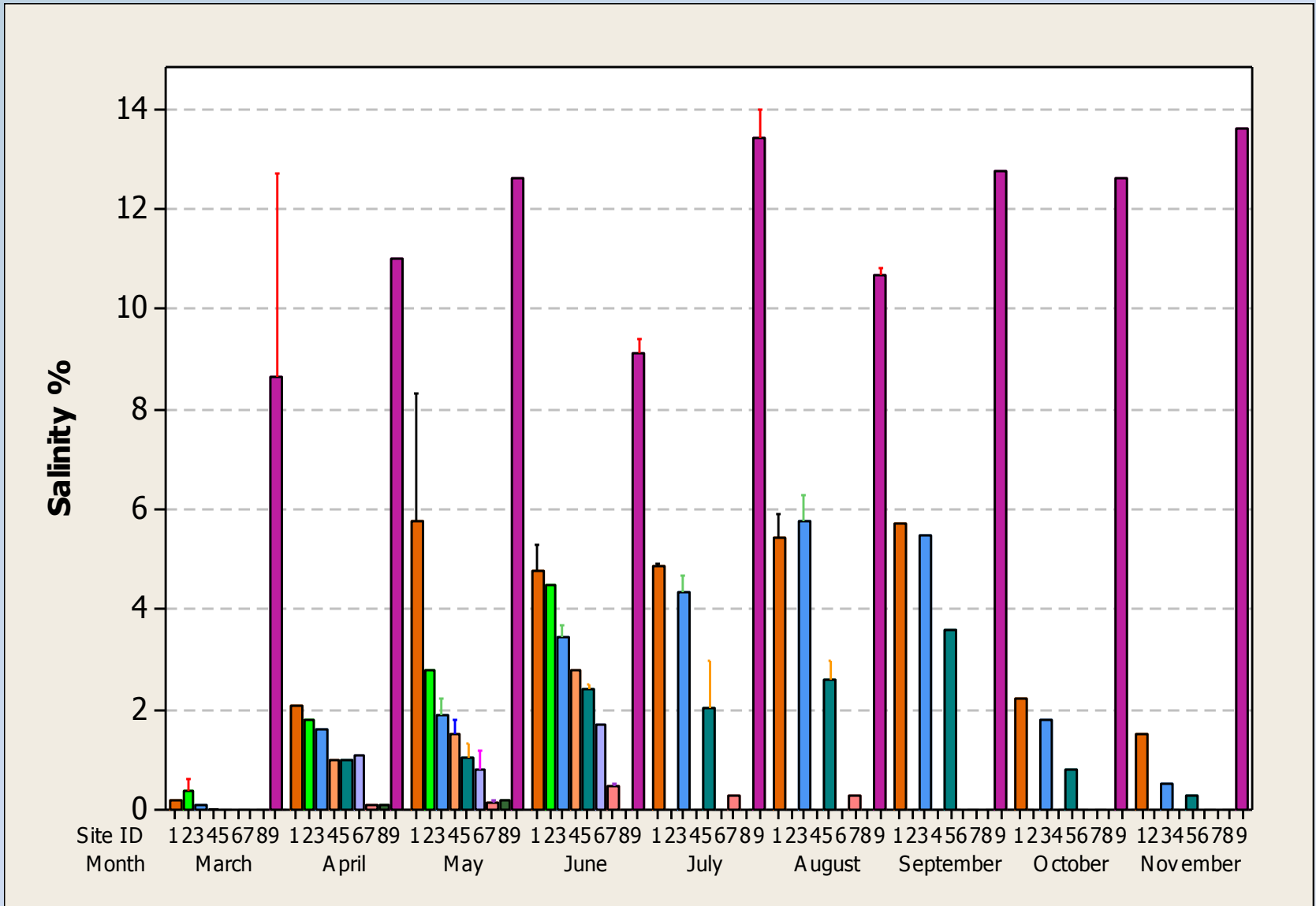
## 14 Transects Surveys

Proposed Date	Actual Date	Proposed Sites	Actual Sites Sampled
Friday, February 15, 2013	Friday, February 15, 2013	2	2
Thursday, March 07, 2013	Thursday, March 14, 2013	9	5
Thursday, April 04, 2013	missed	9	0
Thursday, April 18, 2013	Thursday, April 18, 2013	9	9
Thursday, May 02, 2013	missed	9	0
Thursday, May 16, 2013	Monday, May 13, 2013	9	9
Thursday, May 23, 2013	Thursday, May 30, 2013	9	8
Monday, June 03, 2013	Monday, June 10, 2013	9	8
Friday, June 14, 2013	missed	9	0
Monday, June 24, 2013	Tuesday, June 25, 2013	9	5
Monday, July 01, 2013	Thursday July 11, 2013	9	5
Thursday, July 18, 2013	Monday, July 22, 2013	9	5
Thursday, August 08, 2013	Tuesday, August 06, 2013	9	5
Thursday, August 22, 2013	Monday, August 26, 2013	9	4
Thursday, September 05, 2013	Thursday, September 19, 2013	9	4
Thursday, October 10, 2013	Thursday, October 17, 2013	9	4
Thursday, November 07, 2013	Thursday, November 14, 2013	9	4

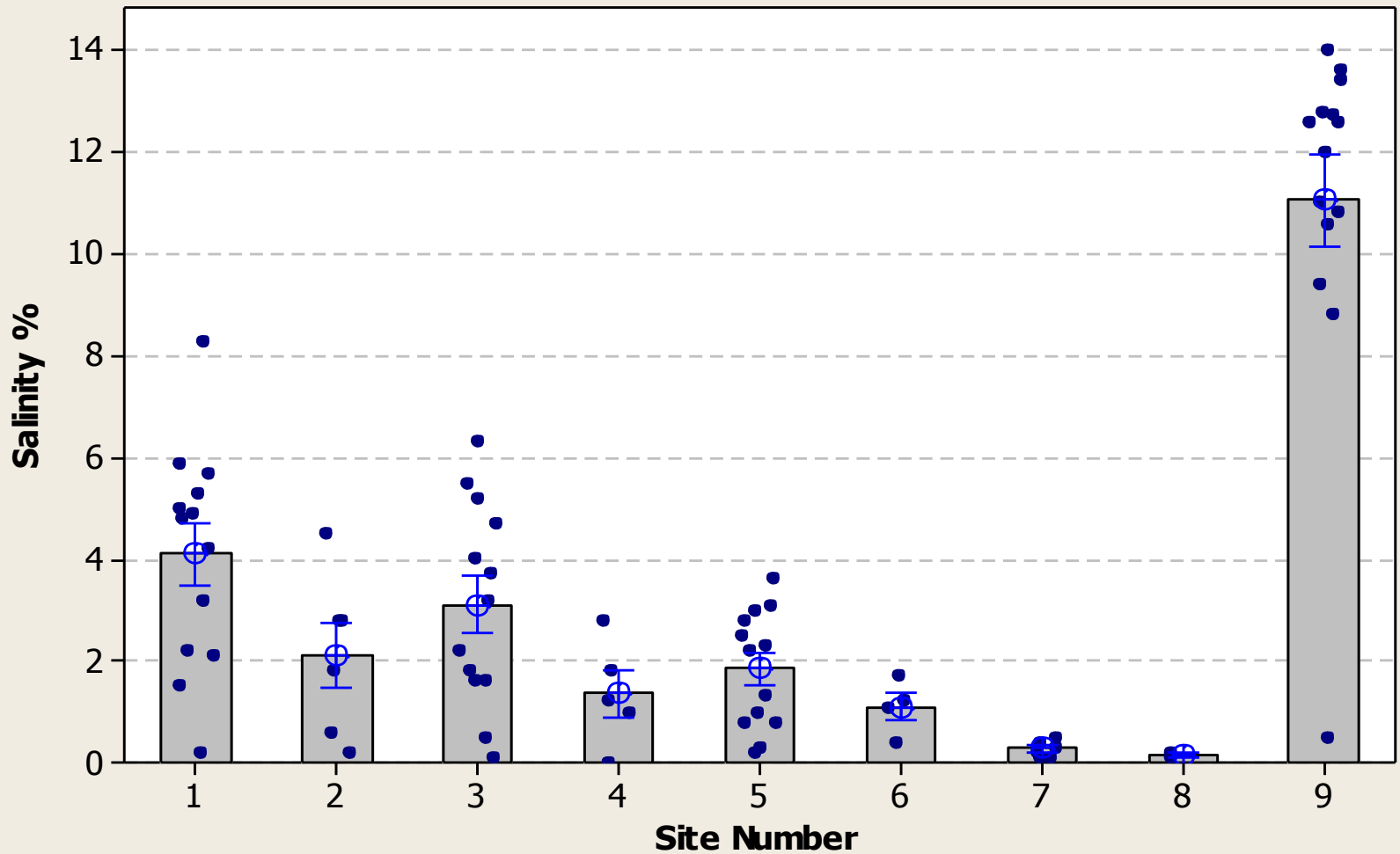




# Temporal Variability of Salinity



# Spatial Variation of Salinity











# Nutrients

## Nutrient Sample Collection and Analysis

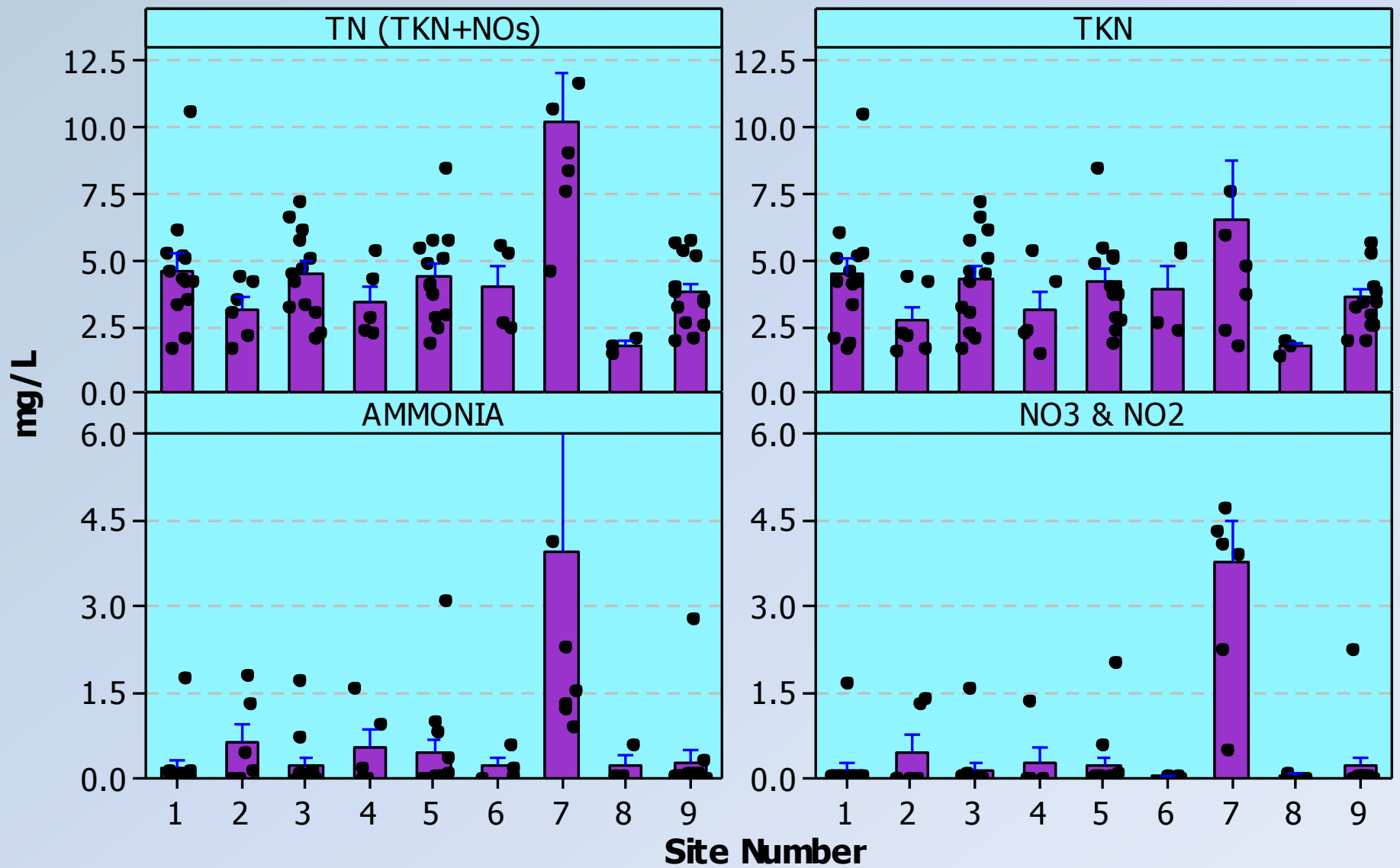
- Ammonia (reporting limit .01 mg/l)
- Nitrate and Nitrite
- Total Phosphorous
- Ortho-Phosphate
- TKN
- Lab: Aquatic research (Seattle)



## Purpose

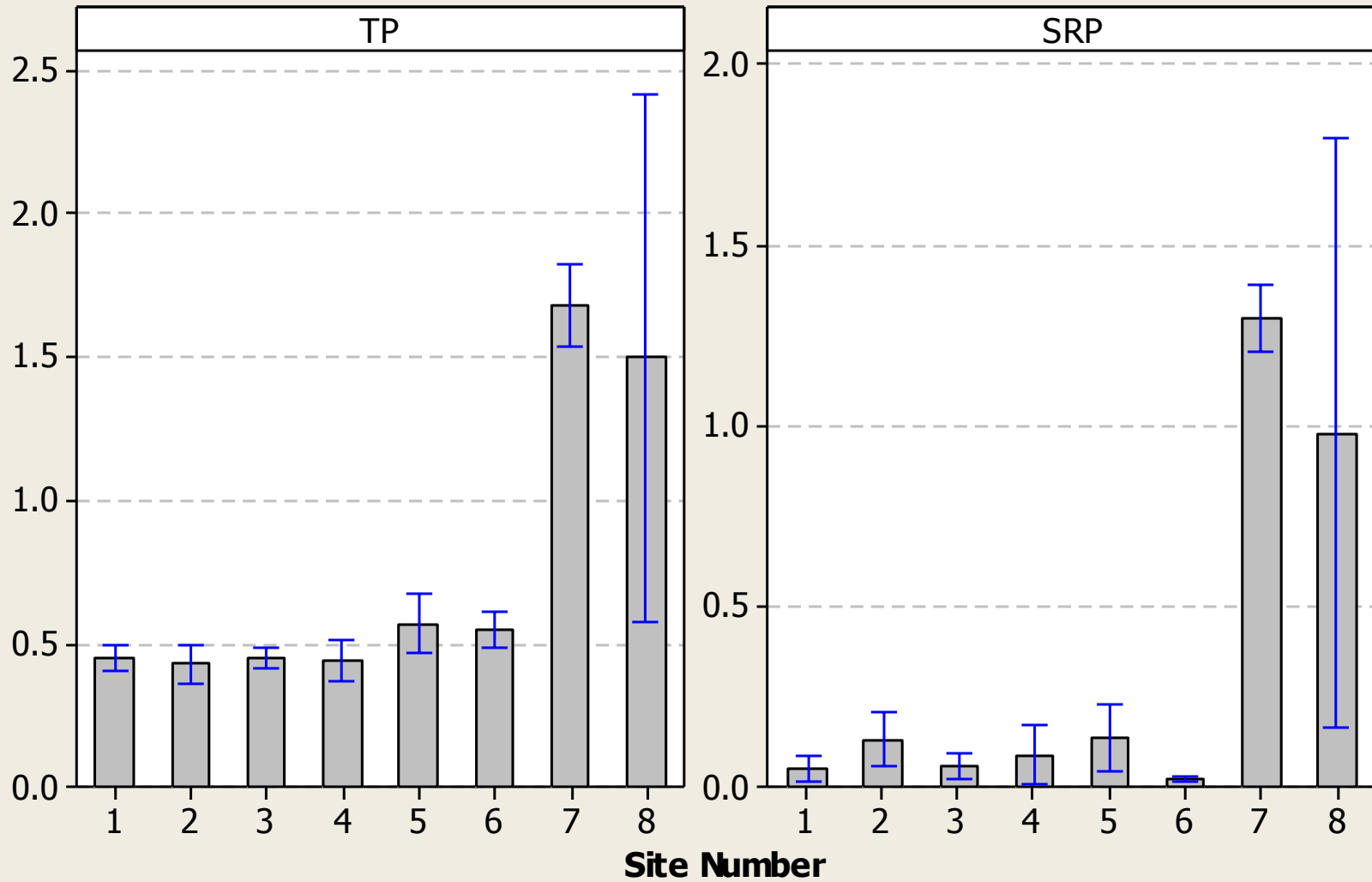
- Evaluate the potential influence that nutrients exert on the algal composition of Farmington Bay.

# Spatial Variation of Nitrogen Species

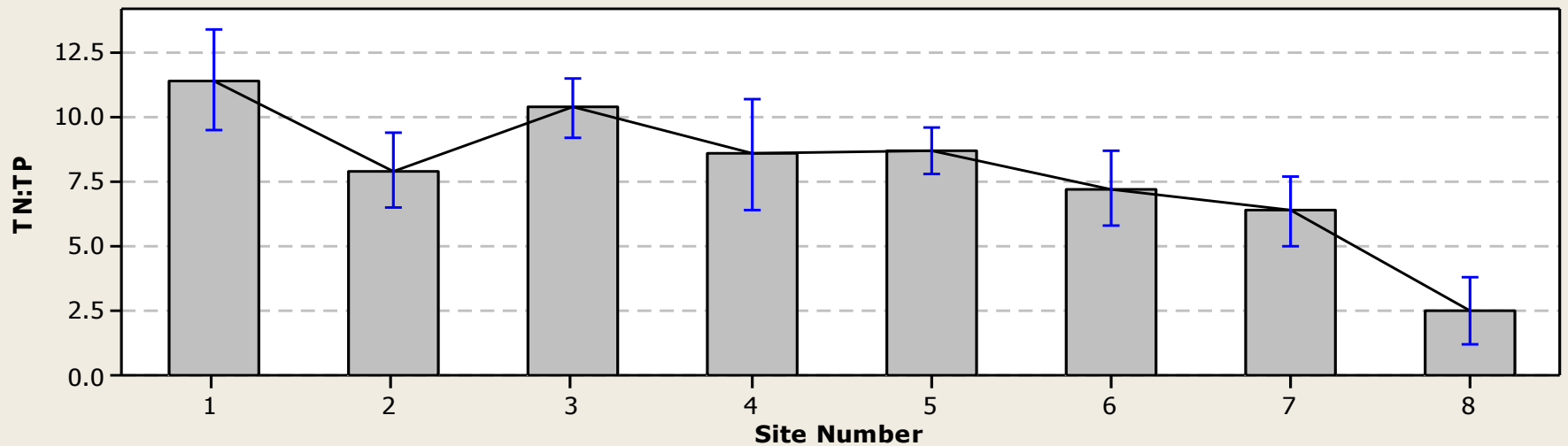
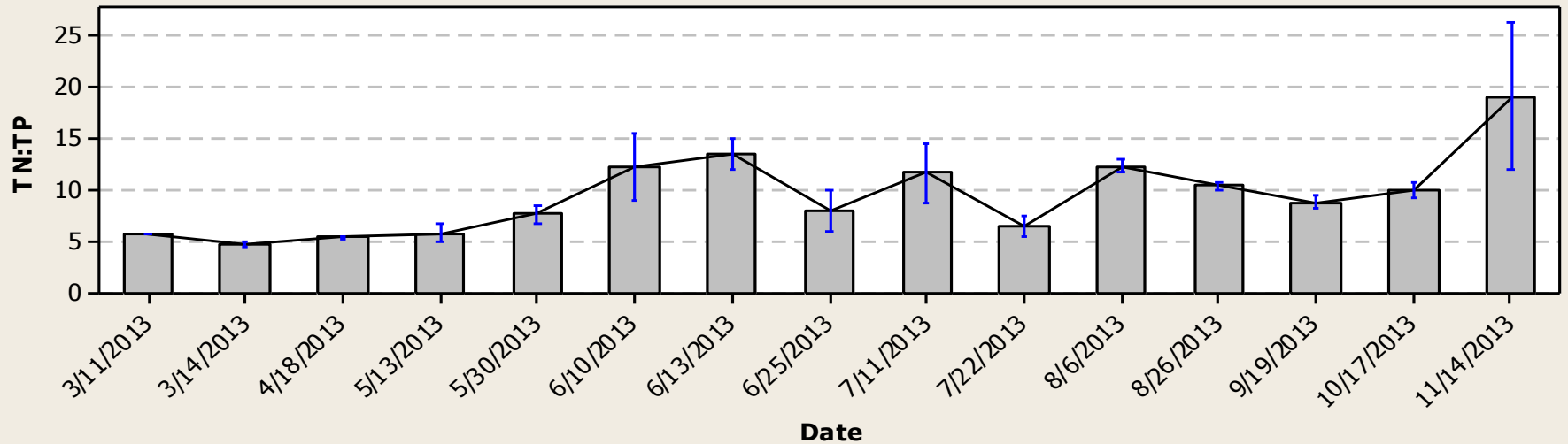




# Spatial Variation of Phosphorus



# Spatial & Temporal N:P Ratio





# Cyanobacteria and Cyanotoxin Analytical Laboratory

**GreenWater**  
laboratories

**Cyano**  
LAB

Tel: 386-328-0882

TF: 877-869-2542

info@greenwaterlab.com

AQUATIC ANALYSIS, RESEARCH, & CONSULTING

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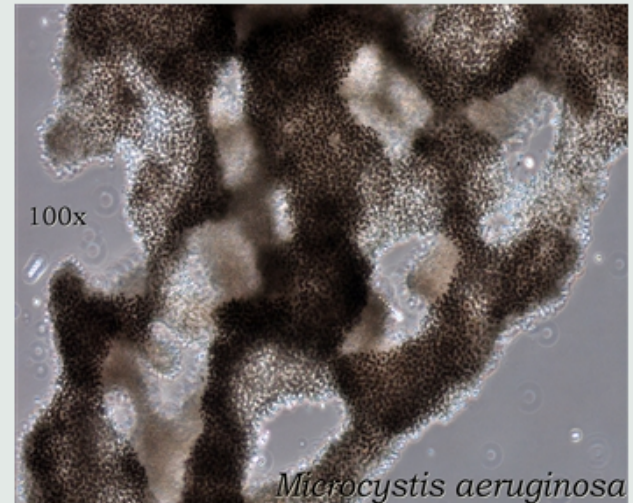
CONTACT US



## Protecting Our Most Precious Resource Through:

- Algal Identification
- Algal Toxin Analysis
- Algal Toxin Standards
- Water Quality Monitoring

Like us on Facebook! 



The background of the slide features a scenic landscape with a calm body of water in the foreground, reflecting the sky. In the distance, a range of mountains with patches of snow is visible under a clear blue sky. The overall color palette is light and serene, with various shades of blue and white.

# Preliminary Results

## Primary Algal Groups

- Blue-green algae: Cyanobacteria
- Green algae: Chlorophyta
- Diatoms: Bacillariophyta
- Cryptomonads: Cryptophyta
- Dinoflagellates: Pyrrhophyta



# Preliminary Results

## Diatoms (Bacillariophyta)



Fig. 4 *Nitzschia* sp. 400X (scale bar = 10μm)

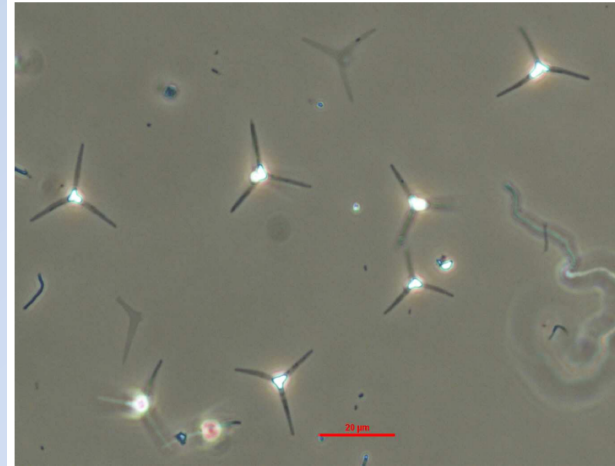
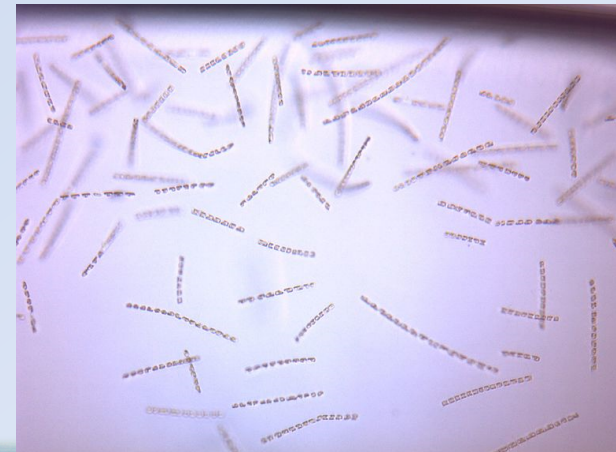


Fig. 1 *Phaedactylum tricordatum* 400X (scale bar = 20μm)



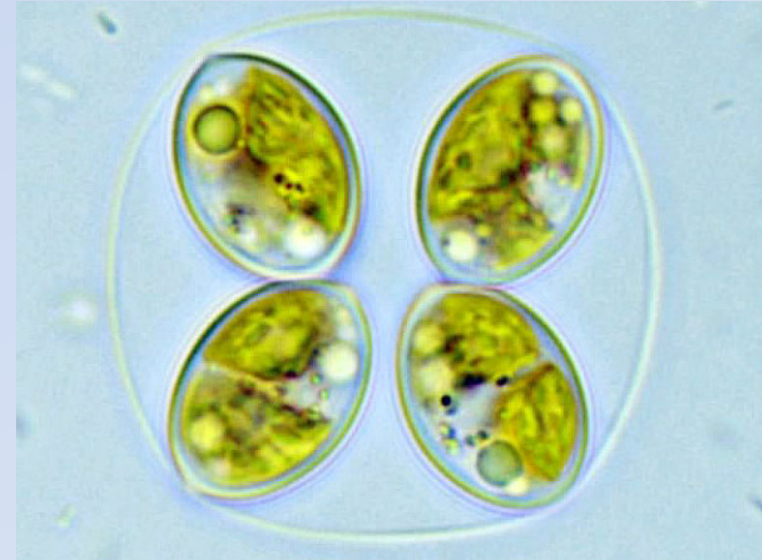
Chaetoceros

# Preliminary Results

## Green Algae: Chlorophyta



Fig. 3 chlorophyte unicells 400X (scale bar = 10μm)



Oocystis

# Preliminary Results

## Primary Cyanobacteria Found in Farmington Bay

- *Nodularia spumigena*
- *Pseudanabena catenata*
- *Pseudanabena sp.*
- *Phormidium sp.*
- *Planktothrix sp.*
- *Spirulina sp.*



# Preliminary Results

## Filamentous Cyanobacteria



Fig. 3 *Nodularia spumigena* 400X (scale bar = 20 $\mu$ m)

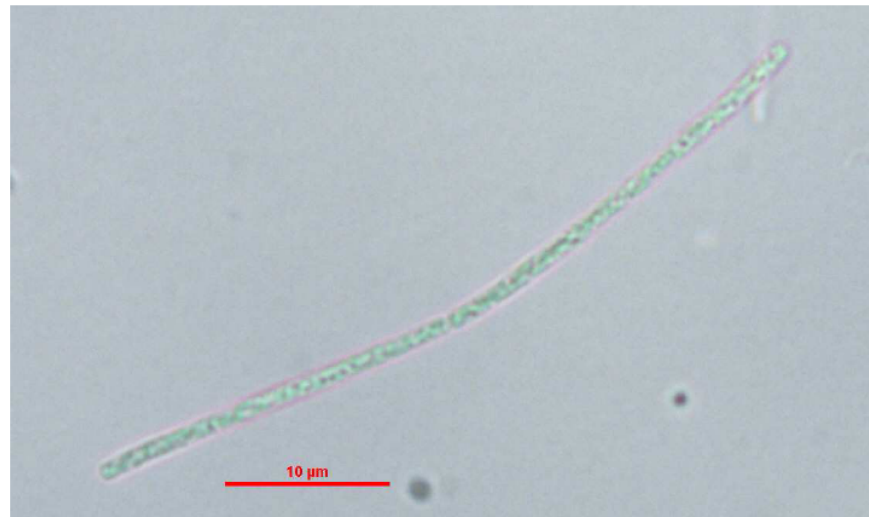


Fig. 1 cyanophyte filament 1000X (scale bar = 10 $\mu$ m)



Fig. 5 *Planktothrix cf. suspensa* 400X (scale bar = 10 $\mu$ m)

# Preliminary Results

## Cyanobacteria: Pseudanabaena



Fig. 2 *Pseudanabaena catenata* 400X (scale bar = 10 $\mu$ m)

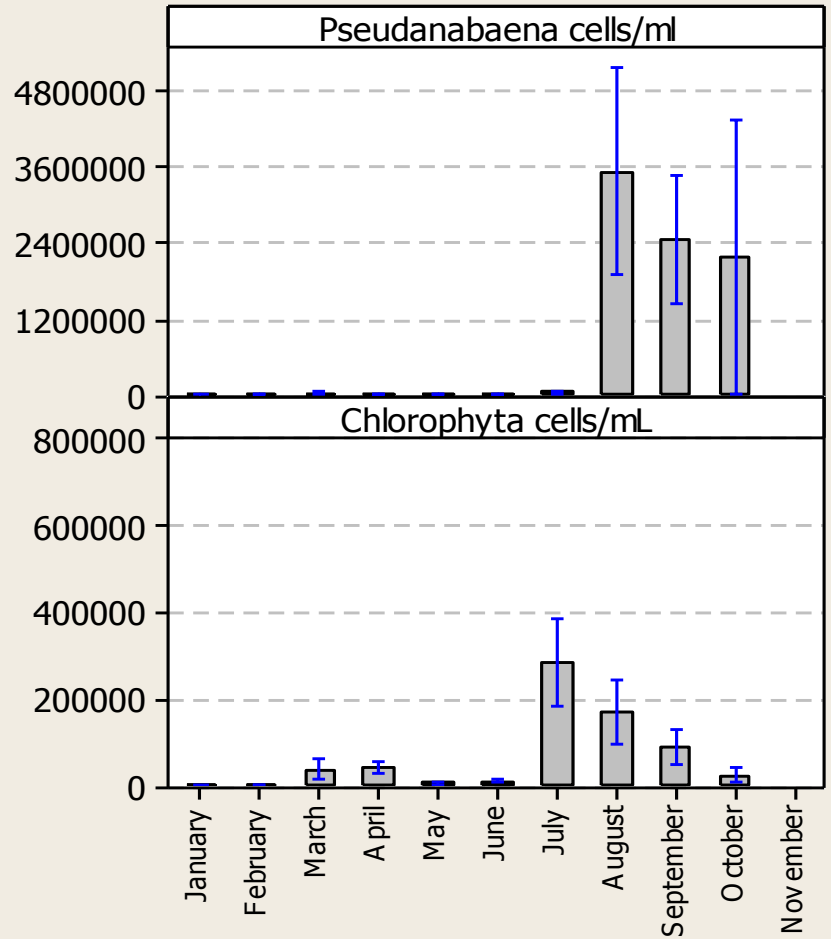
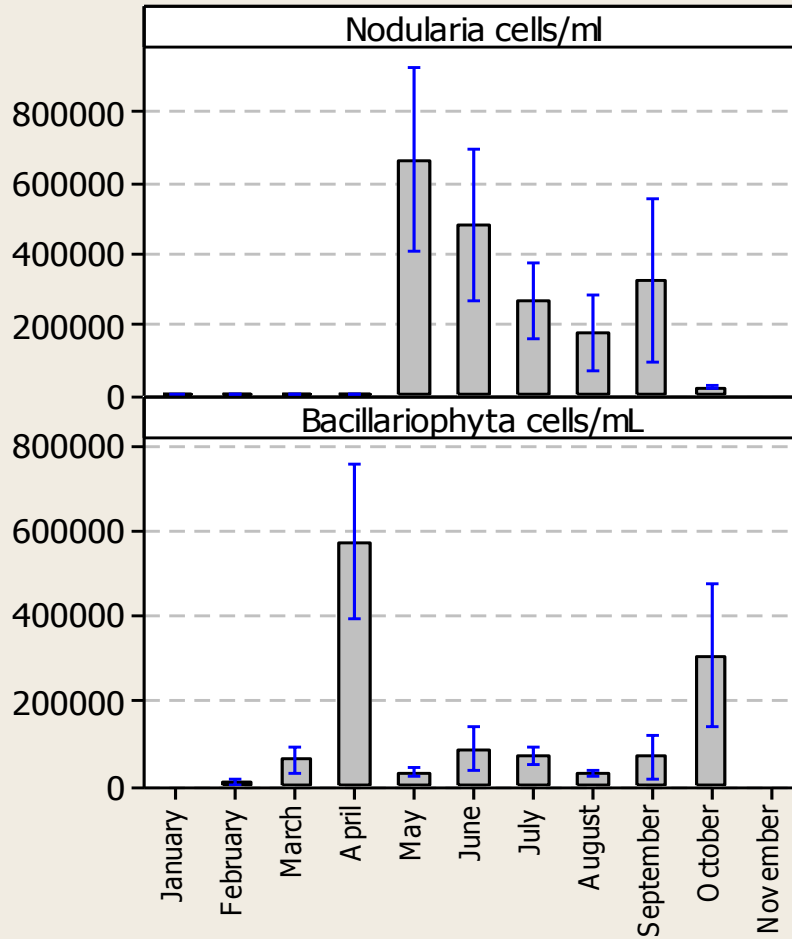


Fig. 4 *Pseudanabaena* sp. 400X (scale bar = 10 $\mu$ m)





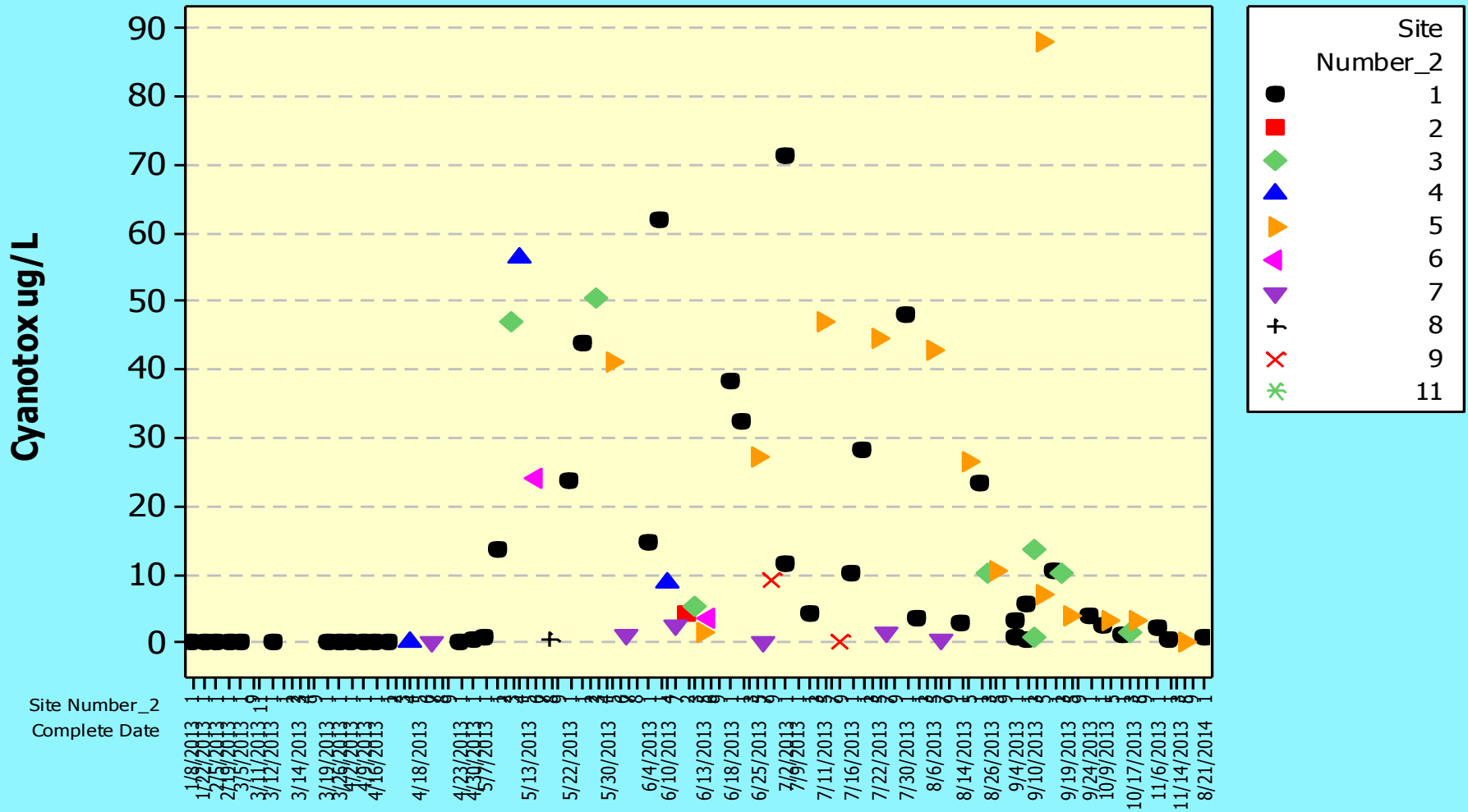
# Temporal Variability of Algae/BG Algae



**Month**

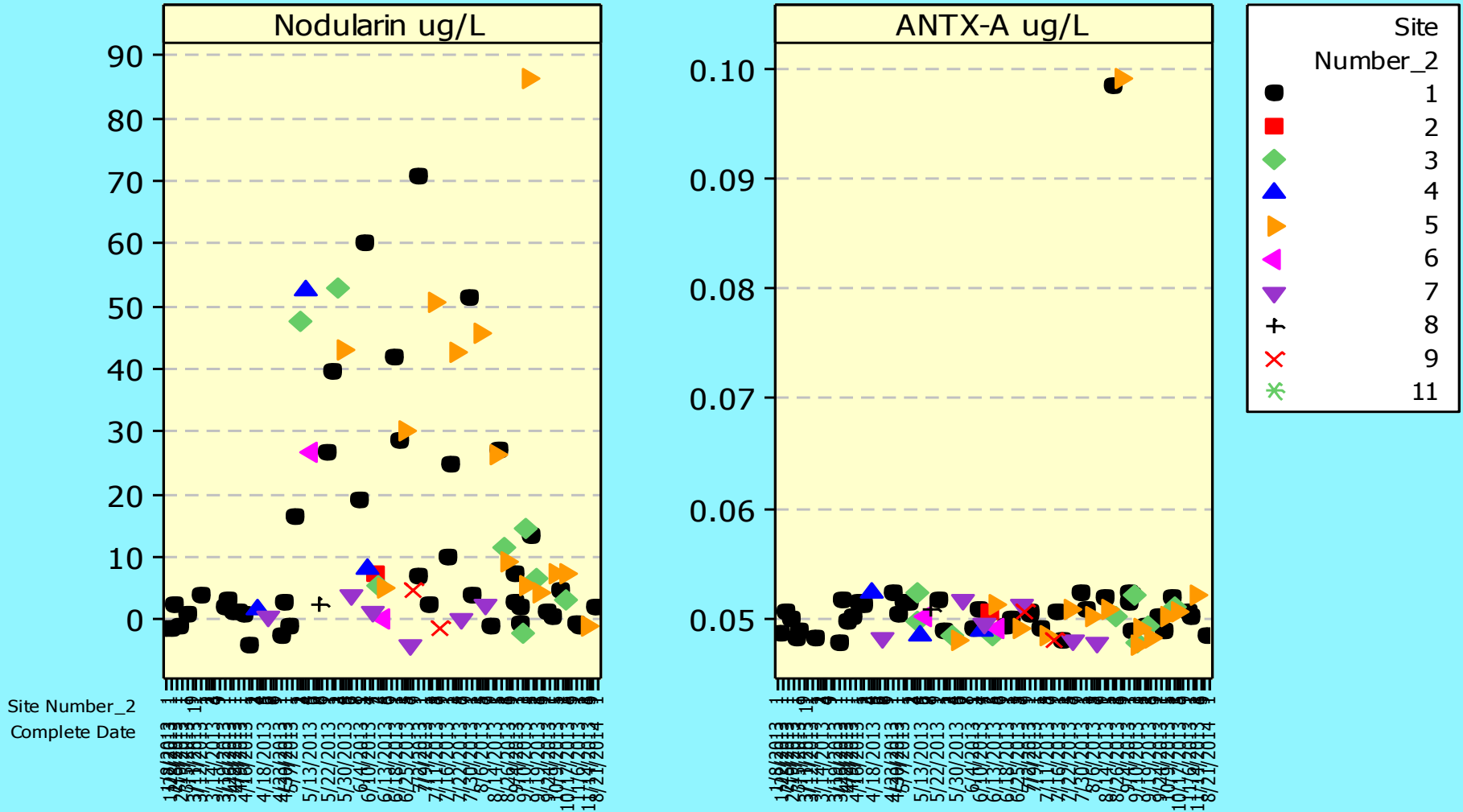
# Preliminary Results

## Total Cyanotoxin Concentration



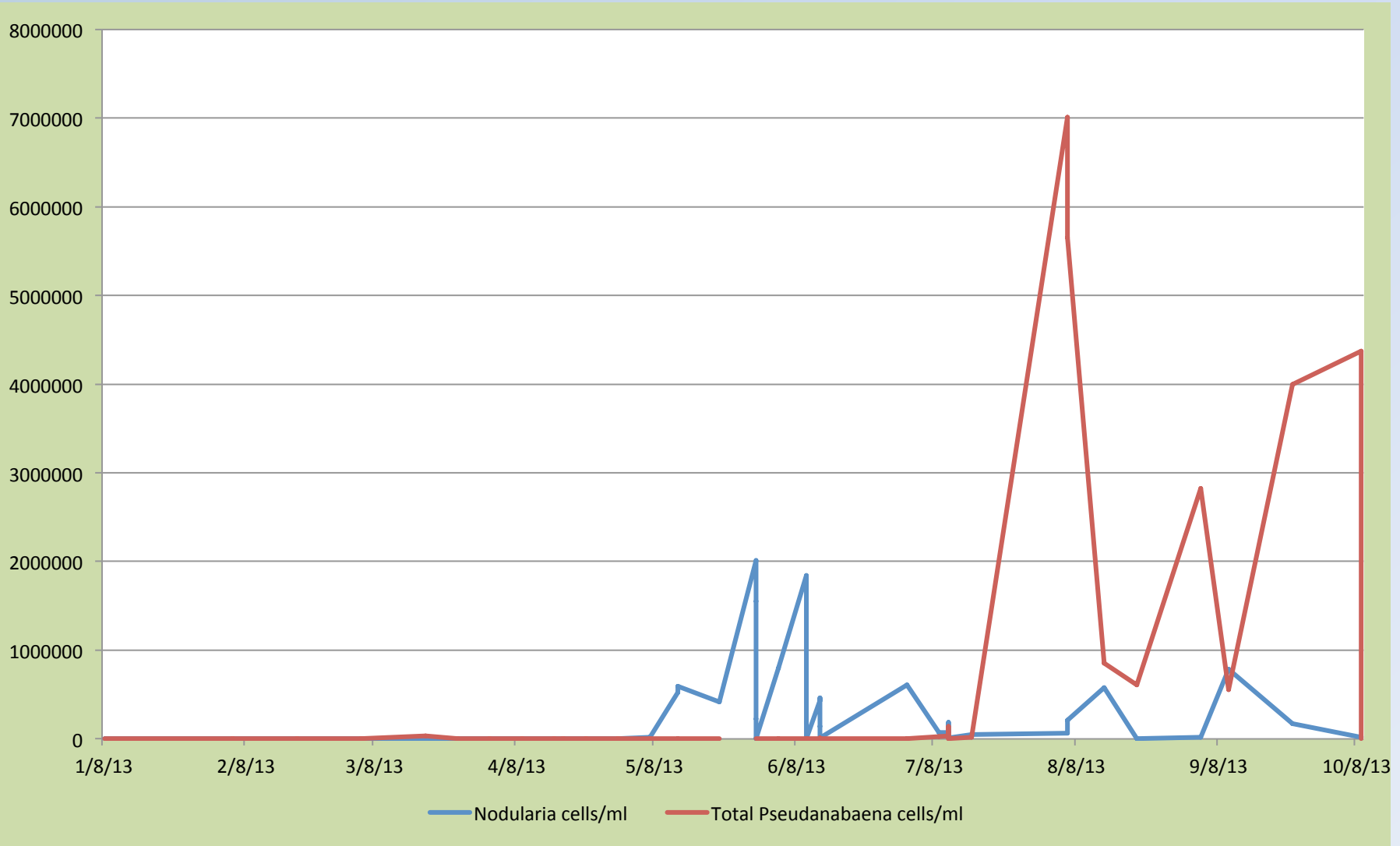
# Preliminary Results

## Cyanotoxins: Nodularin and Anatoxin-a

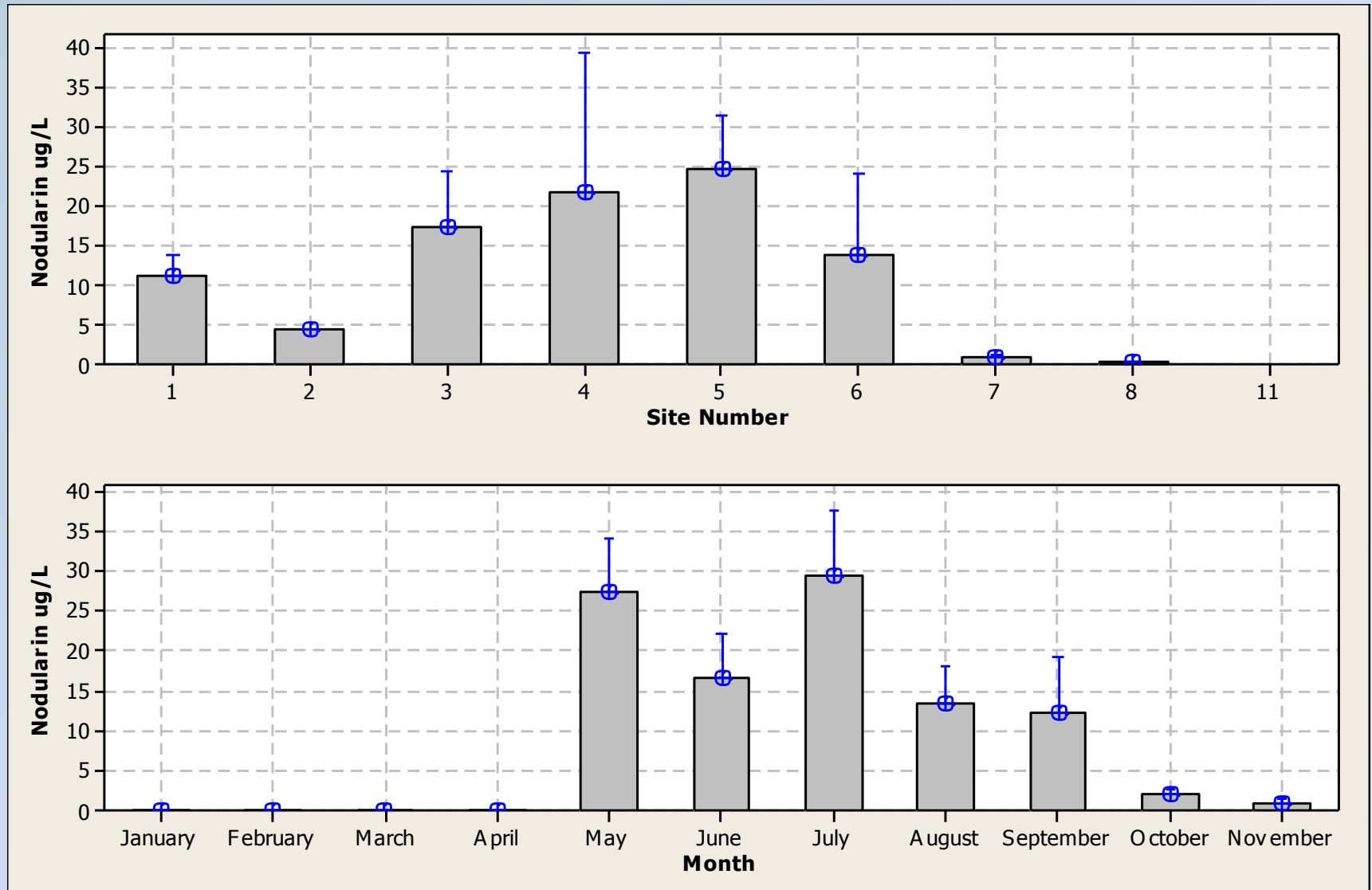




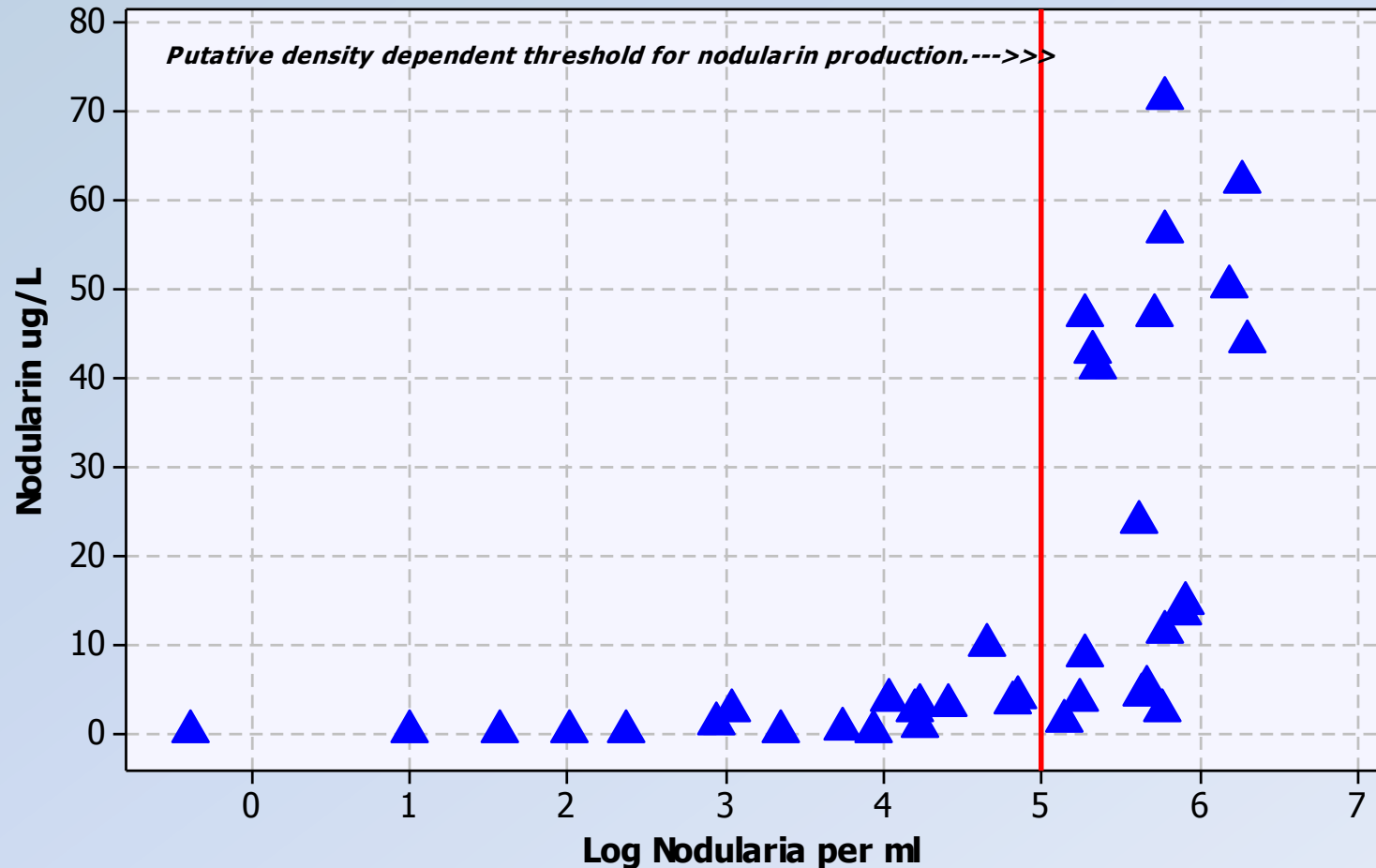
# Nodularia and Pseudanabaena Timing



# Spatial & Temporal Variation - Nodularin



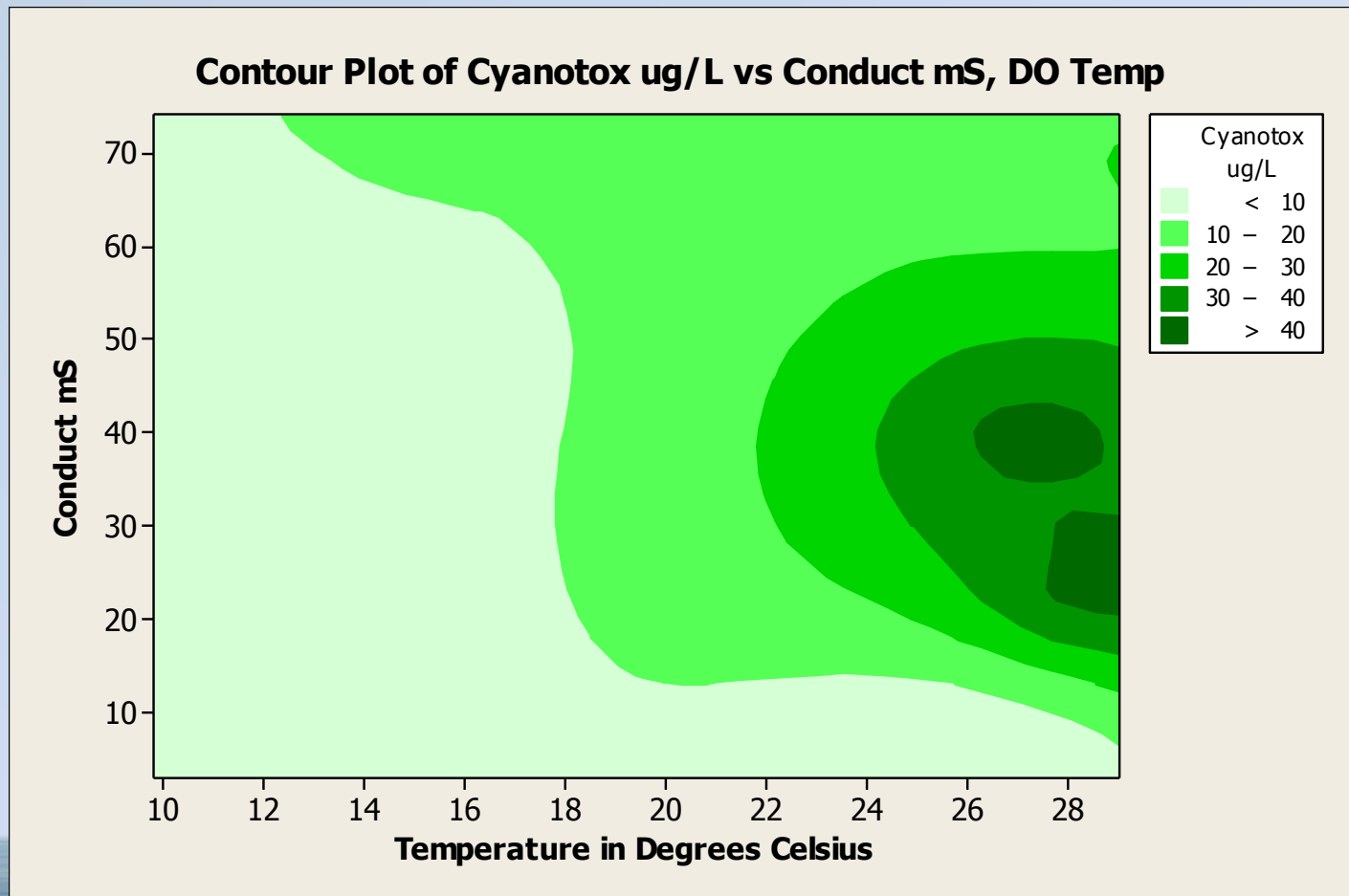
***Nodularia does not uniformly produce the toxin. Rather toxin production appears to be a function of a particular density threshold.***



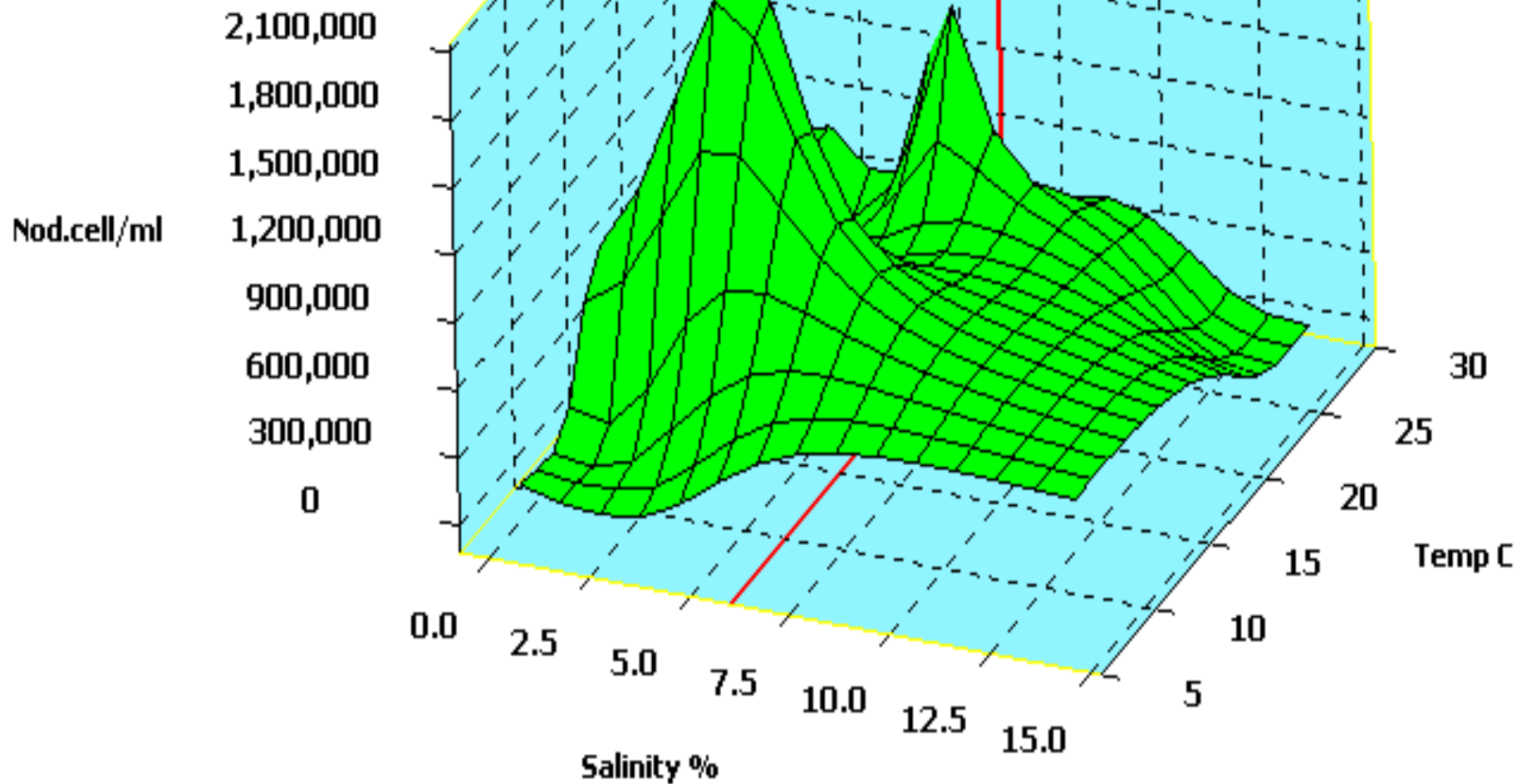


# Preliminary Results

## Comparison of Abiotic Factors and Cyanotoxin Concentration

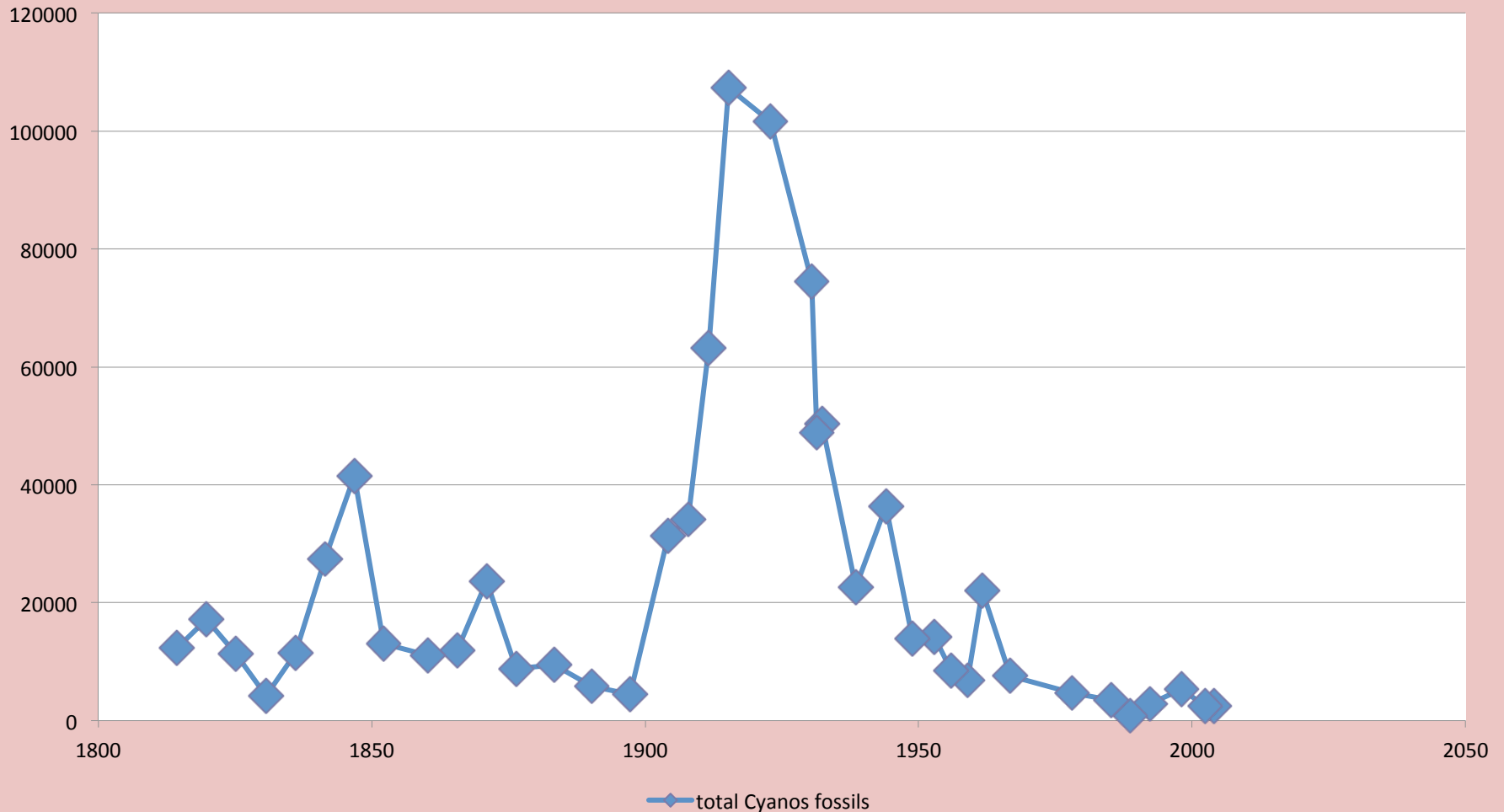


# Nodularia Vs. Salinity & Temperature



# Historic Cyanobacteria Fossil Presence

Total Cyanobacteria Fossils

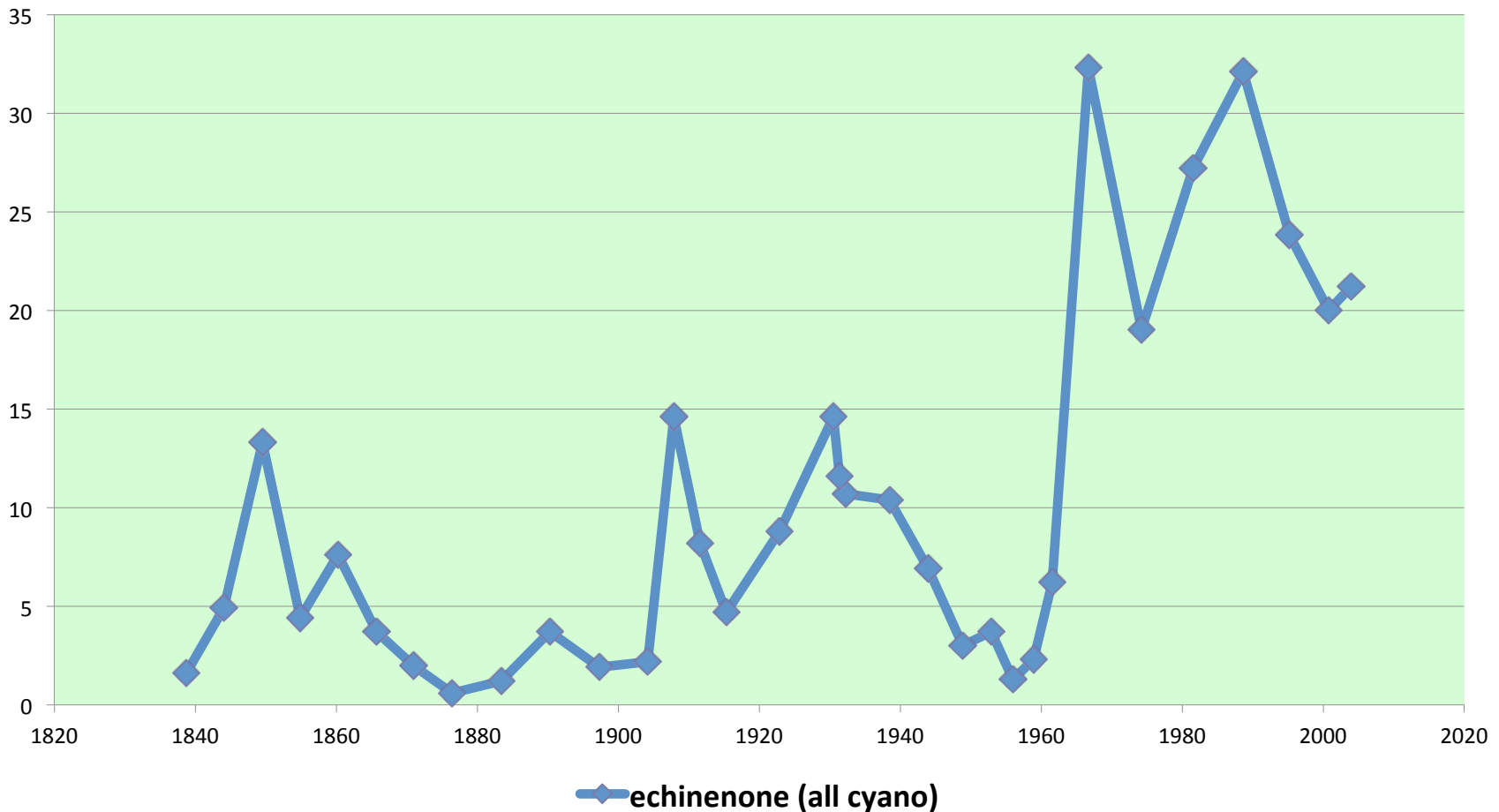


Source: DWQ Paleo Study



# Historic Cyanobacteria Pigment Presence

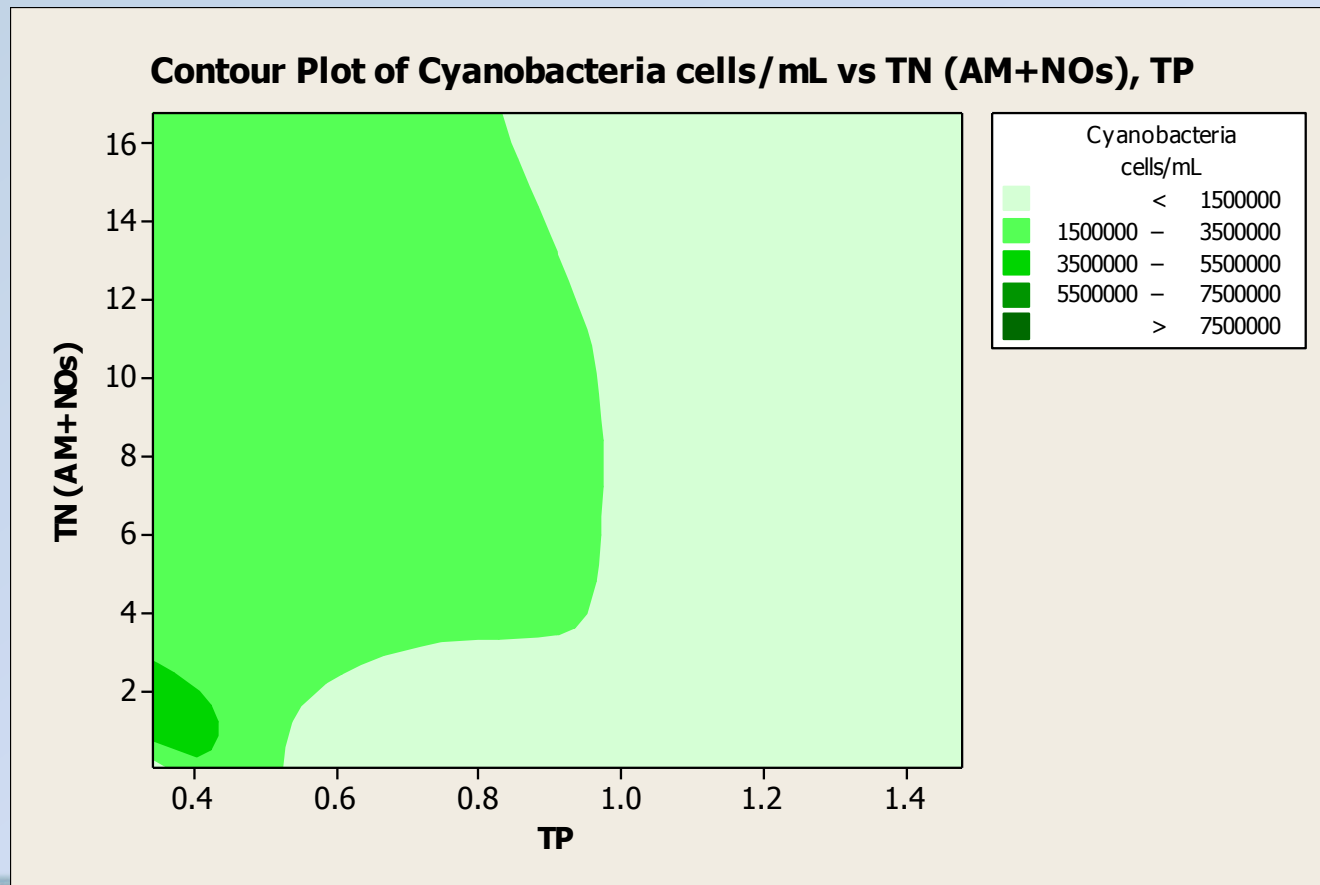
## echinenone (all cyano)



# Preliminary Results

## Algal Composition

### Comparison of Nutrient Dynamics and Cyanobacteria Abundance



# Results

## Zooplankton

### Species Diversity and Population Structure

7 Major Taxonomic Groups

11 Families or Orders

20+ species

77 samples enumerated

Documented very interesting spatial distribution patterns

Collected additional samples for isotopic evaluation

Tracked population dynamics





# ZOOPLANKTON TAXA

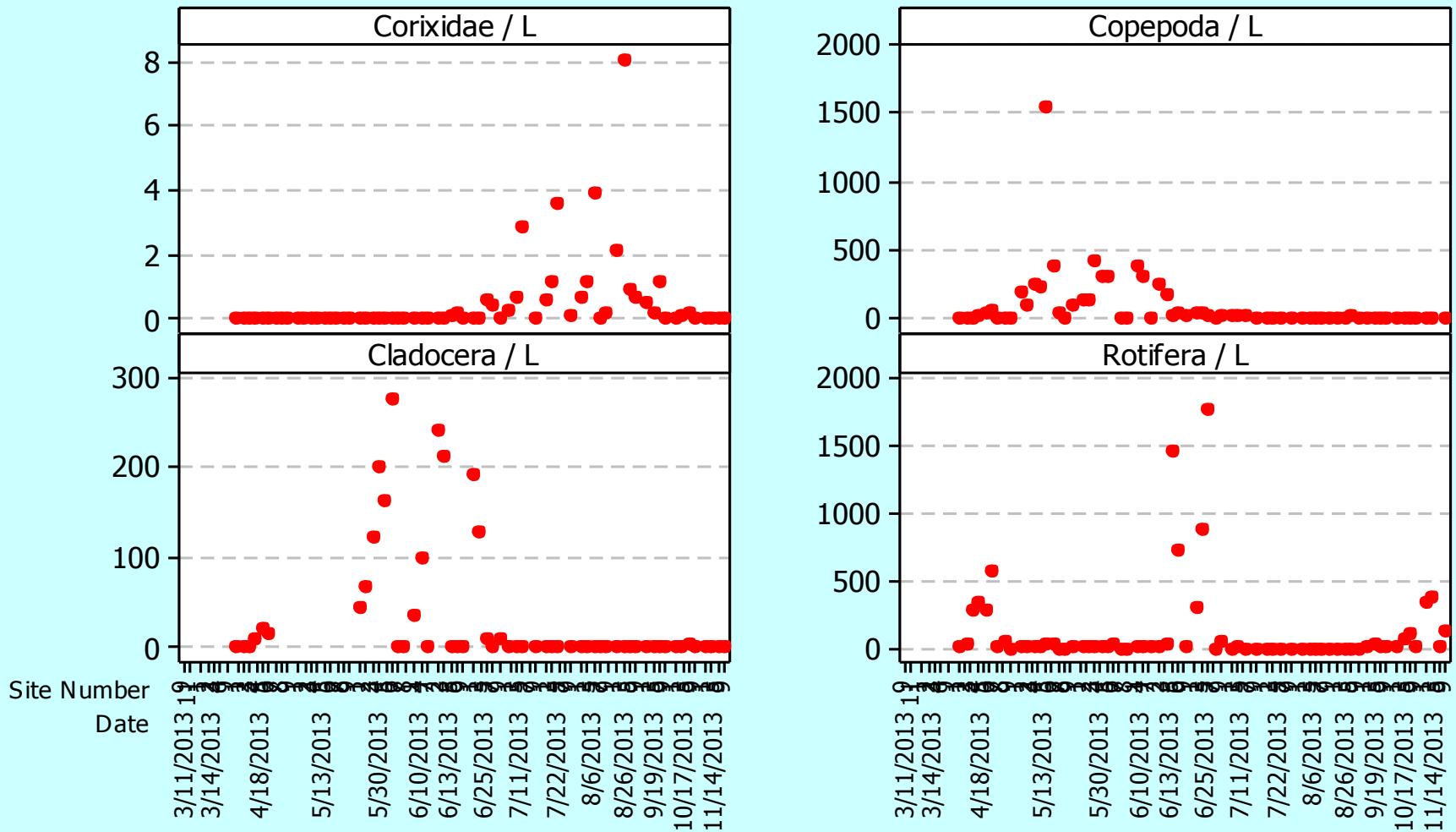
April-November 2013

Species List: GSL Invertebrates from plankton net samples

Major Group (Crustacea)	Family or Order	Species
Cladocera	Daphniidae	<i>Daphnia dentifera</i> (Sars)
Cladocera	Daphniidae	<i>Daphnia pulex</i> Leydig
Cladocera	Daphniidae	<i>Simocephalus vetulus</i> (O.F.M.)
Cladocera	Daphniidae	<i>Ceriodaphniaquadrangula</i> (O.F.M.)
Cladocera	Moinidae	<i>Moina macrocarpa</i> Straus
Cladocera	Chydoridae	<i>Pleuroxusstriatus</i> Schoedler
Cladocera	Chydoridae	<i>Pleuroxus</i> sp.
Cladocera	Chydoridae	<i>Chydorus sphaericus</i> (O.F.M.)
Cladocera	Chydoridae	<i>Alona</i> sp.
Copepoda	Cyclopidae	<i>Eucyclops agilis</i> (Koch)
Copepoda	Diaptomidae	<i>Leptodiaptomus connexus</i> Light
Copepoda	Harpacticoida	<i>Cletocamptus</i> sp.
Branchiopoda	Artemiidae	<i>Artemia franciscana</i> Kellogg
Ostracoda		(undetermined)
Phylum Rotifera	Brachionidae	<i>Brachionusplicatilis</i> O.F.M.
Phylum Rotifera	Brachionidae	<i>Notholcaacuminata</i> Ehrenberg
Insecta: Diptera	Chironomidae	various
Insecta: Diptera	Ephydriidae	(undetermined)
Insecta: Hemiptera	Corixidae	<i>Trichocorixaverticilis</i> (Fieber)
Insecta: Hemiptera	Corixidae	<i>Coriselladecolor</i> (Uhler)
Annelida: Oligochaeta	Naididae	(undetermined)

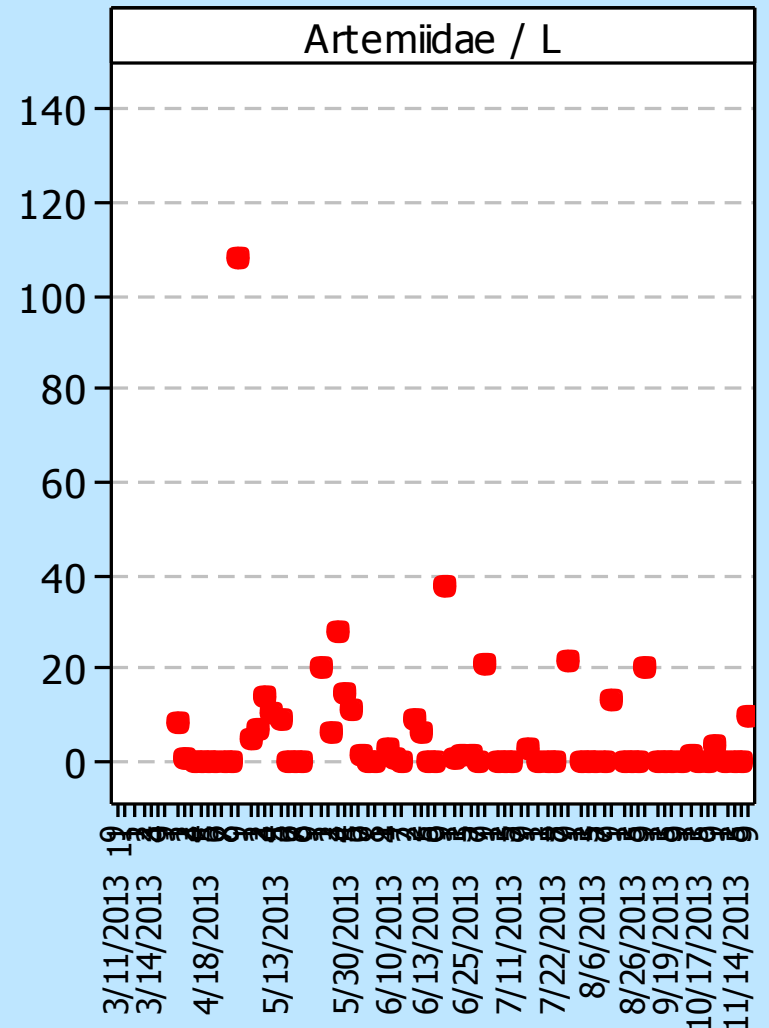
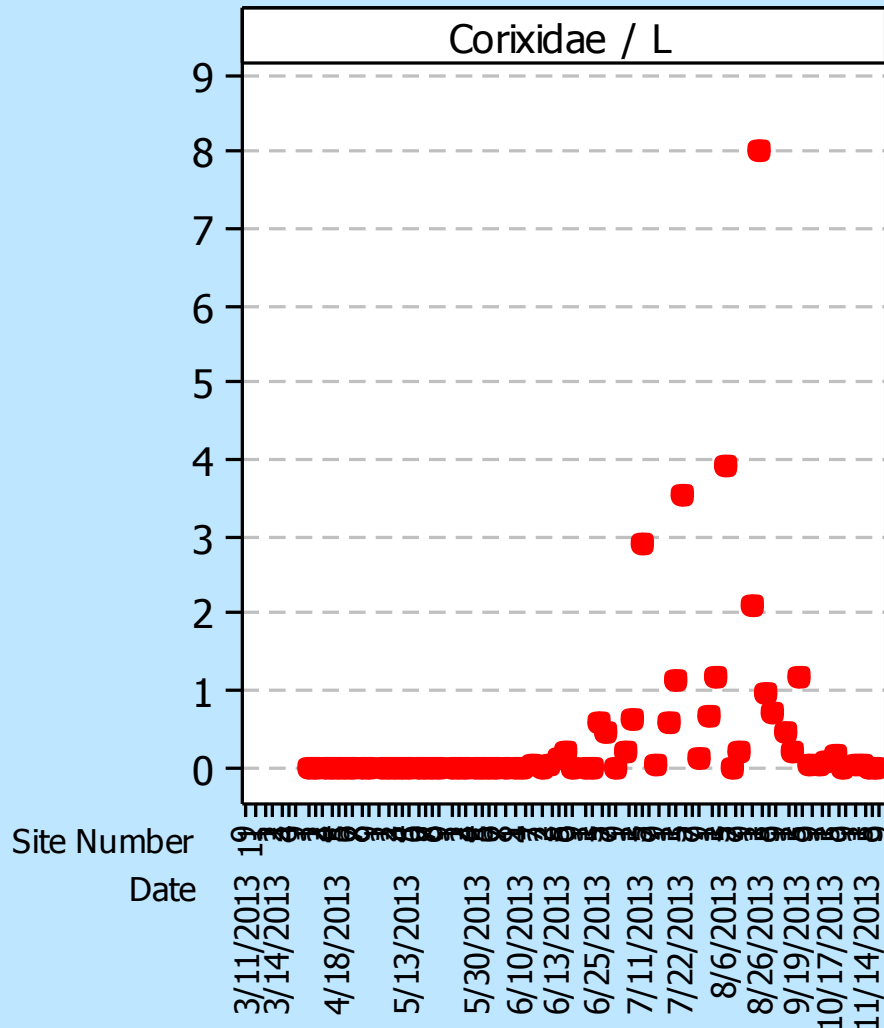
# Comparison of Corixidae, Cladocera, Copepoda, and Rotifera Abundance

## Temporal Pattern of Corixidae and Prey Abundance



# Comparison of Corixidae and Artemiidae Abundance

## Comparison of Corixidae and Artemiidae Abundance





FBAY 7  
6-25-13  
SITE  
1

FBAY 7  
6-25-13  
SITE  
3

FBAY 7  
6-25-13  
SITE  
5

FBAY 7  
6-25-13  
SITE  
7

FBAY 7  
6-25-13  
SITE  
9



FB6

6-13-13

SITE 9



QORPAK® BOTTLES  
BAY # 8  
7-11-2013  
SITE 1

QORPAK® BOTTLES  
BAY # 8  
7-11-2013  
SITE 3

QORPAK® BOTTLES  
BAY # 8  
7-11-2013  
SITE 5

QORPAK® BOTTLES  
BAY # 8  
7-11-2013  
SITE 7

QORPAK® BOTTLES  
BAY # 8  
7-11-2013  
SITE 9

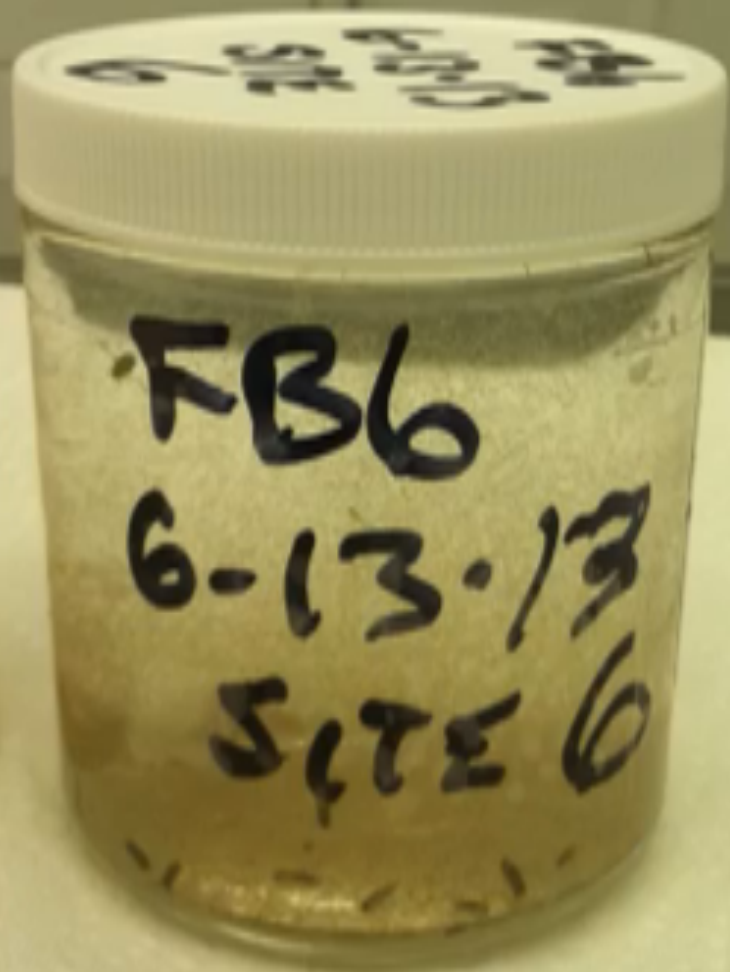
www.essvial.com  
Richmond, VA  
(800) 233-8425



# JULY CORIXID FEEDING FRENZY



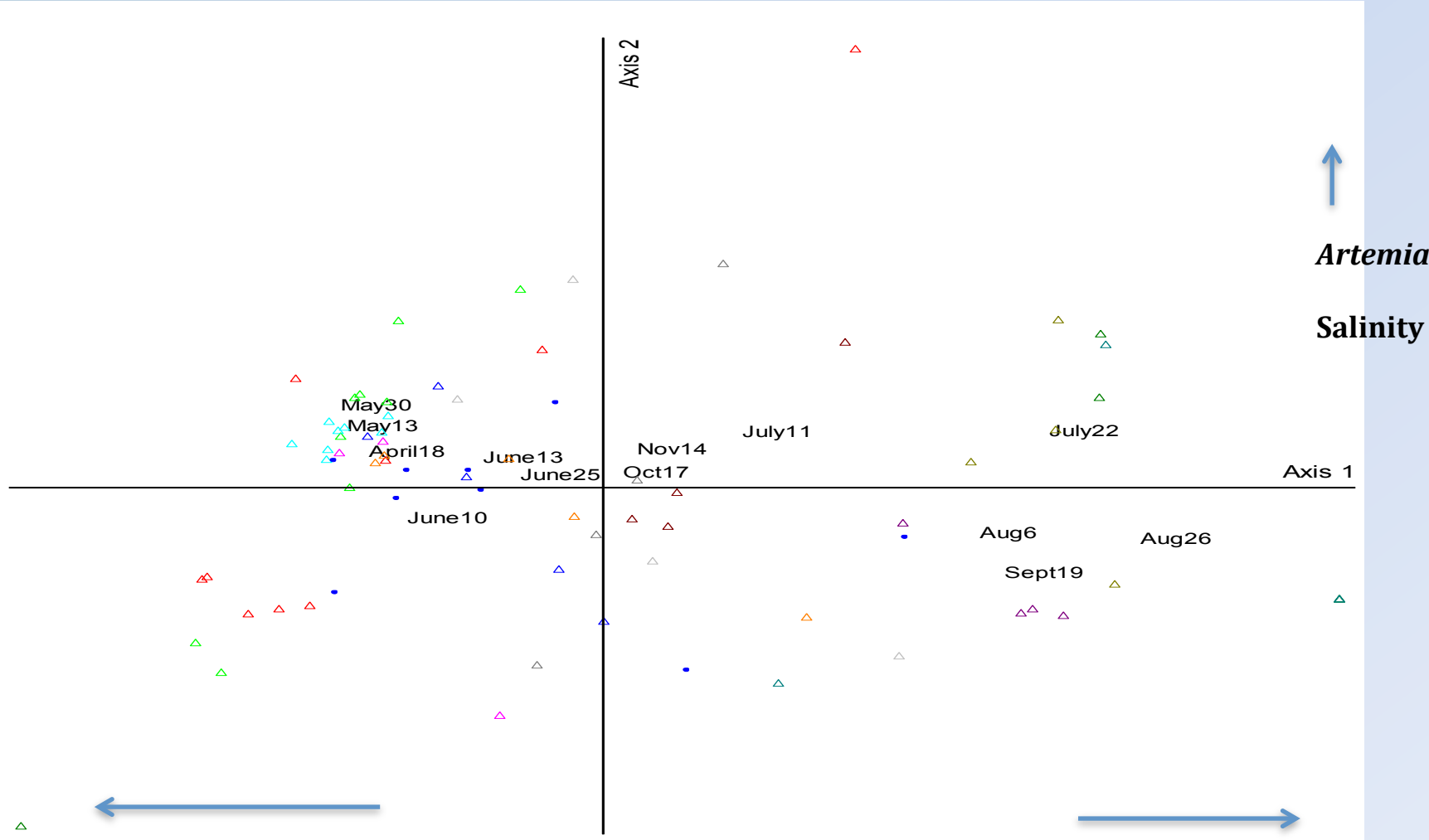




FB6  
6-13-13  
SITE 6

FB6  
6-13-13  
SITE 6

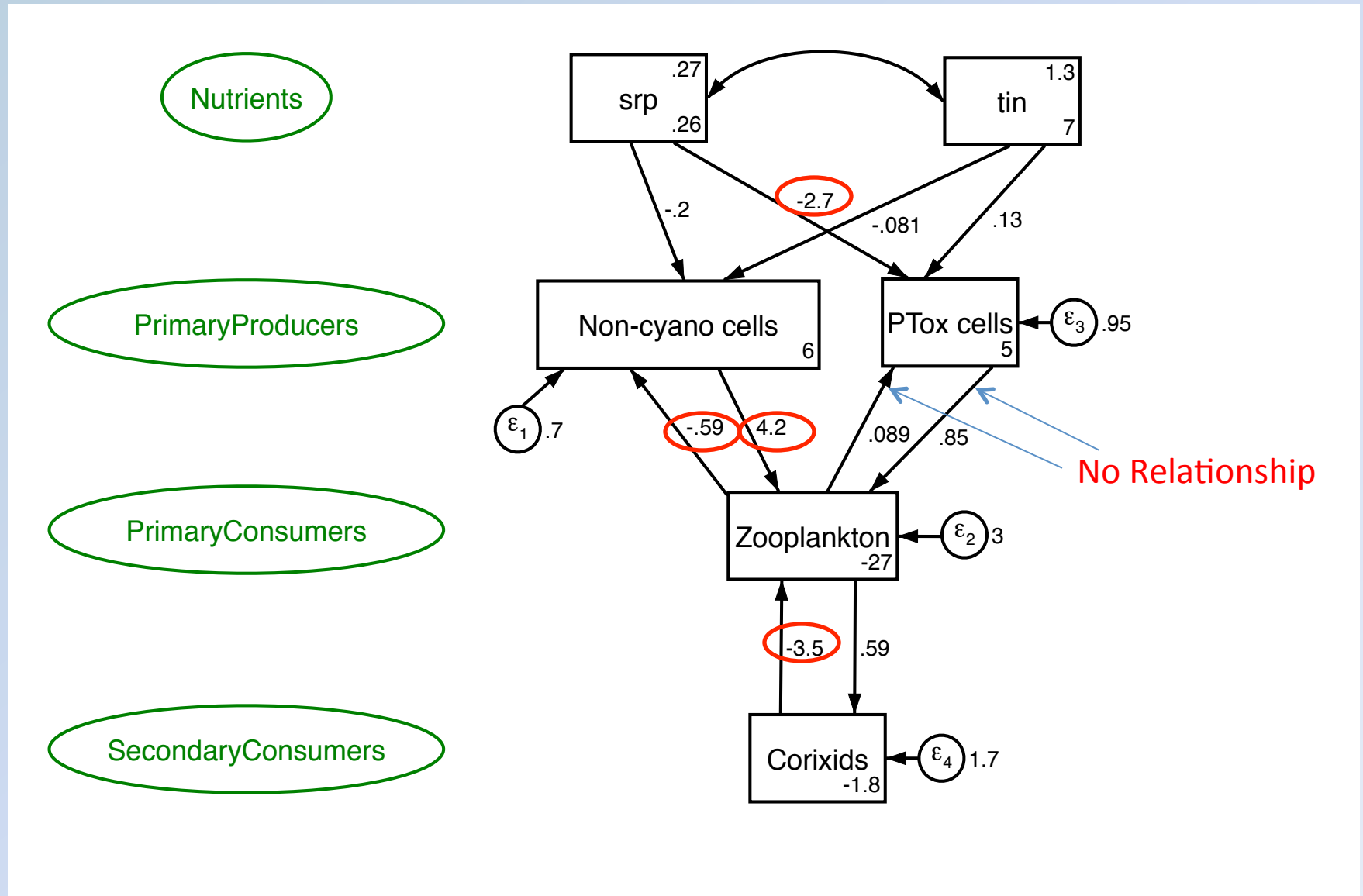
# Non-Metric Multidimensional Scaling Ordination



*Brachionus, Moina,  
Leptodiptomus, Cletocamptus*

*Trichocorixa  
Salinity*

Structured Equation Model illustrating food web relationships in Farmington Bay. Numbers circled in red are significant at  $P < 0.05$ .





# Toxicity Testing

- Dried cysts with known hatching quality are acquired
- Cysts are then suspended in a hatching solution consisting of FBAY water or solution with known concentration of cyanotoxin.
- The suspension is used for 24- to 48-hour hatching using a micro-method—96-well hatching plates in incubator/agitator.
- Recordings of shell content, early breaking, emergent hatching, and nauplii are all recorded.
- Hatching percentage and nauplii survival are recorded.

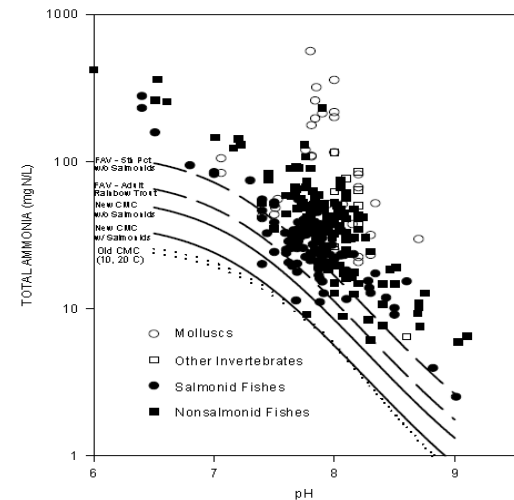
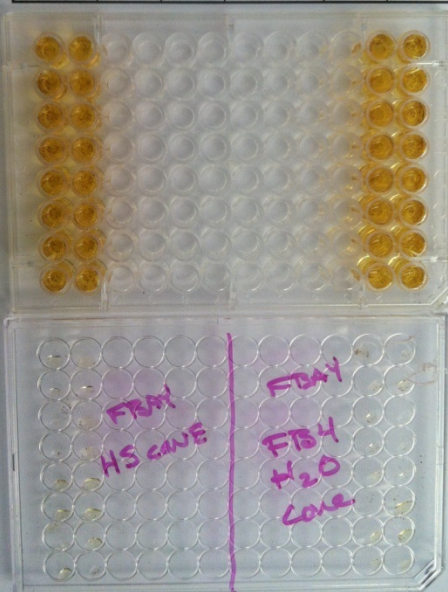


# Exploratory Projects

- Corixid predation Rates
- Stable isotopes in algae and zooplankton
- Water quality impacts on hatching and survival of Artemia nauplii

Initial Artemia Toxicity Testing: Microplate Bioassay Method

METHOD	SOLUTION	Nauplii	umbrella/cysts	% nauplii	Average
cone	hatch sol	315	37	89.5%	89.6%
cone	hatch sol	316	36	89.8%	
cone	fbay #4	314	40	88.7%	
cone	fbay #4	317	26	92.4%	90.6%
microplate	hatch sol	252	51	83.2%	
microplate	hatch sol	259	50	83.8%	83.5%
microplate	fbay #4	254	76	77.0%	
microplate	fbay #4	264	36	88.0%	



# And then there was Utah Lake

October 5 & 6, 2014 – Two Dead Dogs



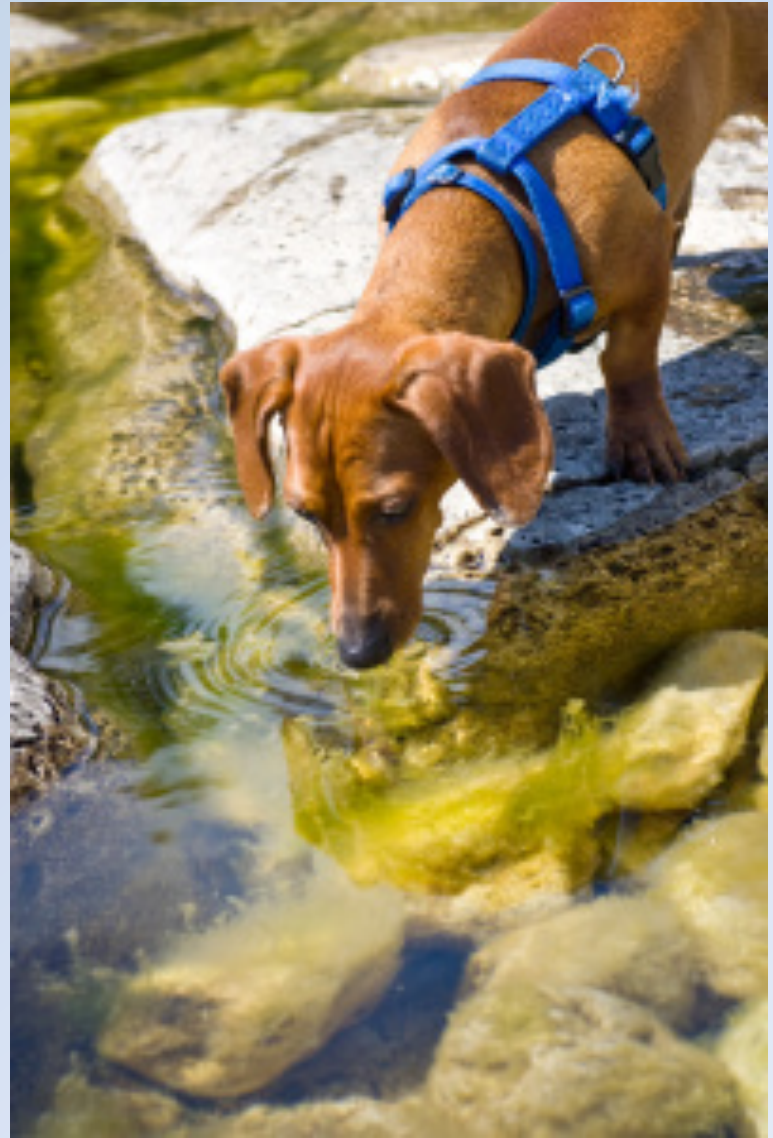


# Canary in the Mine



=





# Measured Toxin Concentrations

Microcystin:      Max 11 ug/L  
                              (WHO 20 ug/L)

Anatoxin            Max 0.2 ug/L  
                              (lowest Limit 1 ug/L)

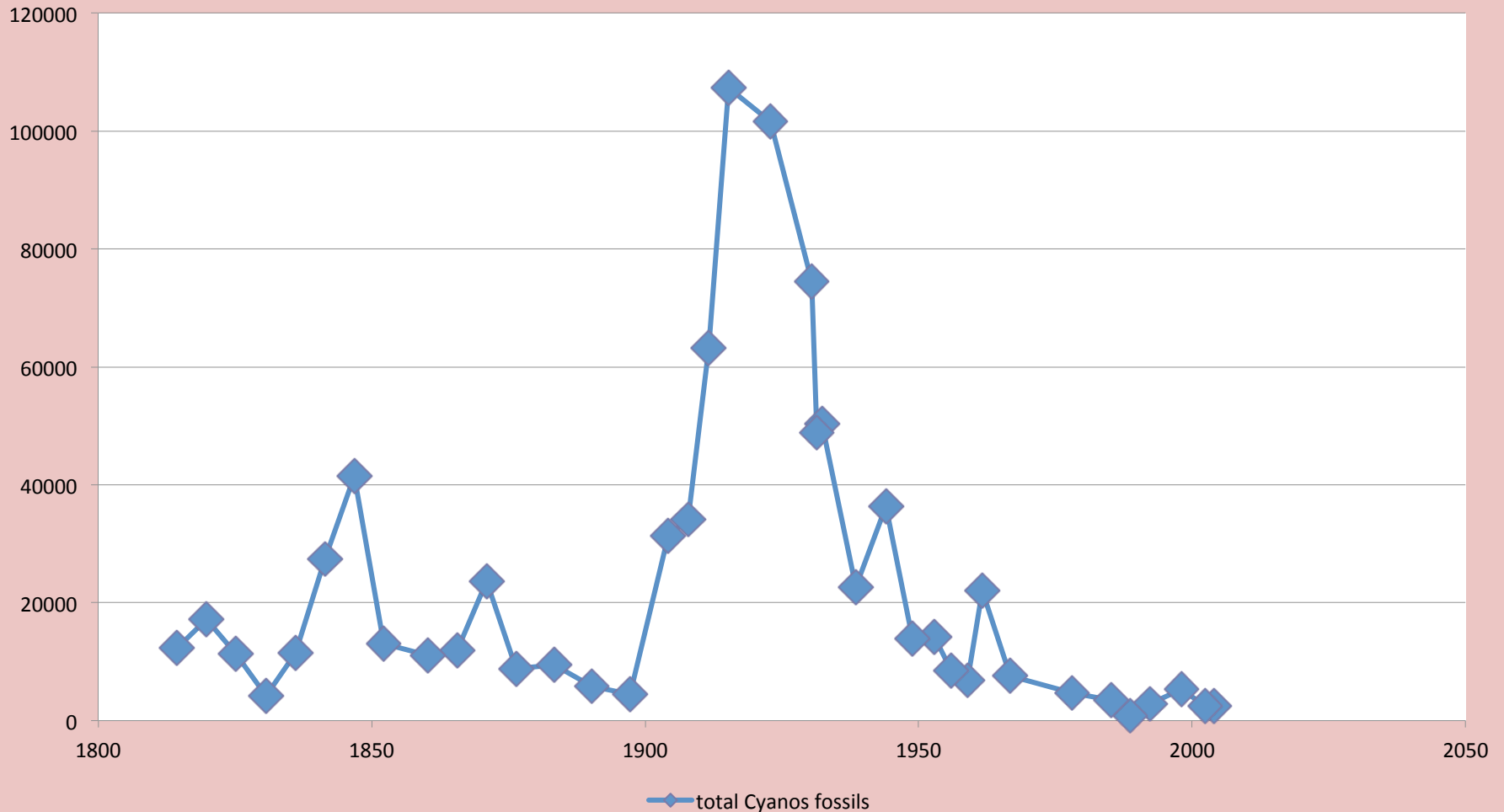


# UTAH DWQ

The preliminary necropsy report for the dog that died shortly after drinking the water on October 5, 2014, from Utah Lake has been finalized. A specific cause of death was not identified. The Utah Veterinary Diagnostic Laboratory concluded that cyanobacteria toxins were an unlikely cause of death because no cyanobacteria or cyanotoxins were detected in the stomach contents or liver of the animal.

# Historic Cyanobacteria Fossil Presence

Total Cyanobacteria Fossils



Source: DWQ Paleo Study

# Anatoxin Detection Problems

There are problems with anatoxin-a detection methods including an inability to determine whether it is anatoxin-a or another neurotoxin that causes the resulting deaths.

**So, what do we do?**



# ALGAE BLOOM ADVISORY

Algae blooms have been spotted in this water body.

- Don't swim, wade or fish near blooms or surface scum.
- Don't drink the water.
- Keep children and animals away from any blooms or surface scum.
- Rinse with clean water if exposed.



For more information please visit our website at [www.nyc.gov/parks](http://www.nyc.gov/parks).

City of New York



Parks & Recreation

# Maintain a Neutral and Objective Perspective



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**Questions?**